

REVIEW

Unflinching boon of Nanotechnology on Plant Growth

Garima Tripathi, Soumyadeep Basu, Shrestha Dutta, Shweta Tripathi*

*Corresponding author. Email : (stripathi@amity.kol.edu)

Department of Biotechnology, Amity University Kolkata, New Town, Kolkata, India 700135

Abstract

In the current paradigm of science, nanotechnology has acquired a remarkable threshold in diversifying its effect on technology. Several approaches have been made towards the frontier of plant growth and enhancement. Nanotechnology proves to be the prime substantial and effective edge in the era of advancing domain. The main prerogative of the article is to showcase the varied prospects of nanotechnology in spite of its certain glitches, which are yet to be tapped in the near future. The amount of knowledge we have received, although being less, has much scope for refinement. To impact the three domains of life - flora, fauna, and mankind, the manifold comprehension of nanotechnology provides humungous opportunity of growth.

Keywords

Nanomaterials, Nanoscience, Microscopy, Spectroscopy, ENPs, Phytotoxicity, Anti-bacterial effect, Nanoherbicides, Nanofertilizers, Biosensor

Introduction

Nanotechnology is the protean as well as integrative area of inquiry and application. The wide range of applications that nanotechnology has to offer and will be providing, speaks of its extensive prevalence, whether it be in the field of agriculture, energy, electronics, medical purpose, healthcare sector, textiles, commuting means, constructions, cosmetics, water treatment etc., nanotechnology has an integral part to offer or rather it has an indispensable role to play.[1-4]

The origin and ideology behind nanoscience and nanotechnology began with a lecture titled, "There's Plenty of Room at the Bottom", by physicist Richard Feynman at an American Physical Society gathering at the California Institute of Technology on 29 December, 1959. Feynman, who is widely recognised for being the Father of Nanotechnology, elucidated nanotechnology to be an exercise in which scientists will be able to manoeuvre and handle individual atoms and molecules. Almost after a decade, Prof. Norio Taniguchi, in his analysis or investigation of ultra accurate machining, conceived the name, "Nanotechnology". It was with the improvement of scanning tunnelling microscope, that the visualisation of individual atoms was made possible and modern nanotechnology started its journey.

Nanoscience is the survey that deals with the occurrence and administration of constituents at atomic, micro and macromolecular levels, in order to comprehend and make use of the properties that differ distinctively from those on a larger picture. This type of extraordinary and diversity unfolds the new prospects for more knowledge and application.

Nanotechnology, being such an inspiring medium of technology of rising techno-economic archetype, is still in its dawning stage of interpretation, advancement and innovation. Keeping in mind its characteristics, the experimentalists have considered this to be an effective tool of sustaining technology for the long run. As defined by the United States Nanotechnology Initiative (2001,) that nano-technology is the understanding and control of the matter and dimension of roughly 1-100 nanometers, where unique phenomena enable novel application. Encompassing the nanoscience, engineering and technology involves imaging, measuring, modelling, and manipulating matter at this length scale.

Nature has numerous evidence of nanotechnology, based on its capability to work at atomic, micro molecular and supra molecular levels. Nature is the ultimatum in raising nanostructures that provide functional proteins and compounds at cellular level of great importance in life on earth. Biological systems have been considered by some scientists to involve separation and sectionalisation of many structures into a stipulated pattern or device. Some biological systems comprise nanosystems that are dedicated towards particular functions such as mobility, function, etc. The preparation of nanomaterials comprise a straight and artificial way that reaps particles in nanosize range, following grinding, high pressure amalgamation and disruption to diminish the size.[5,6] On the contrary, the bottom up process in the synthesis of nanomaterials that deal with reactive

volatile precipitation and solvent displacement.[7] It is very important for deep introspection to understand that nanomaterials due to their enhanced contact surface area might be lethal, an effect which might be absent in its bulk counterpart, mainly in the open agricultural setup.[8,9] Examples include, stable mold with nanomaterials that have nanostructure freely packed to its surface, where it can reasonably be expected to percolate once coming in contact with aqueous medium or air, or when unveiled to rationally anticipated mechanical forces.[10]

The contribution of nanoscience in plant growth and system is acquiring a major importance in the present scientific scenario. Nanomaterial having different physical and chemical attributes can result in improved plant metabolism. Pros and cons of ENPs on the environmental consequences with biological results. Nanoparticles can cause biochemical, physical and physiological changes in plant system and development. Impact of MWCNT and SWCNT on plant architecture, has triggered a scope of in depth study on nanotechnological aspects in plant world. The phytotoxicity, stress, ecological condition are now put in limelight for further research to make nanotechnology a beneficial tool in the improvement of agricultural yield.[11]

2. Categorization of Nanomaterials

Nanomaterials are those particles whose size ranges between 1 to 100nm. The leading types of nanomaterials are— carbonaceous[12], semiconductor, metal oxides[13,14], lipids[5], zero-valent metals[16,17], quantum dots, nanopolymers[18], and dendimers[19] with various types of nanofibres, nanowires and nanosheets.

Nanomaterials have a special physical and chemical propagations, and the ability to enhance plant metabolism. Galbraith et al(2007) and Torney et al(2007) conform that engineered nanomaterials are efficient in effortless transport into leaves, DNA parts and other biomolecules into plant cells[20].

The engineered nanoparticles have been broadly helped in diverse area of science like bioengineering, electronics, textiles, chemical engineering, advancement of green environment, pharmaceutical shipment system in navy machinery. The exclusive asset of ENPs is analysed as broad surface area, large surface energy, and adept quantum restrain.[21,22]

When nanoparticles are disclosed into the environment, ENPs may modify their movement by means of physical, biochemical and biological conversion. It requires great realization that the disadvantages of nanomaterials due to their enhanced contact surface area might be harsh especially in an agro economic set up. Engineered nanoparticles are those particles which contain metals as its main constituent especially metal oxide. The yield of metal oxides and metal nanoparticles could be accomplished by various ways. There are various array of nanoparticulate metal oxides including both single oxides (CeO_2 , TiO_2 , ZnO , CrO_2 , MoO_3 , and Bi_2O_3) and binate oxides (BaTiO_2 , LiCoO_2 , and SnO). This sequence of metal

oxides has modern utilization in UV inhibitory effect and detectable clarity of nanoparticles foam. ZnO and TiO₂ are largely used in cosmetics, sunscreen, and bottle coatings. In the year 2005-2010, it was recorded that ZnO and TiO₂ have dermatological benefits. To augment the grade of emission, CeO₂ is used as a combustion catalyst in fuels like diesel, and also in oxygen pumps, gas sensor, solar cells and metallurgical ceramics.

Nanoparticles have been broadly classified under two headings – Organic and Inorganic Nanoparticles.

2.1. Organic Nanoparticles :

2.1.1. Micelle :-

Micelles are nanostructures composed of amphiphilic molecules like polymers or lipids. [23] When exposed to aqueous environments, they mask their hydrophobic group inside the structure and unveil the hydrophilic groups. Whereas in a lipid rich environment their structure may set up in a reverse way. [23]

2.1.2. Dendimer :-

A dendimer is morphologically identified by a branched structure grown from one or more cores. The size of these NPs is effortlessly controlled by number of generations that are allowed to breed over these cores. Dendimers offer complication regarding drug incorporation and release, being their synthesis quite time consuming. [24]

2.1.3. Liposomes :-

Liposomes are unilamella spherical structures composed of amphiphilic compounds and present high production cost and content leakage. The main advantage of liposome is that they are completely biodegradable, non-pernicious and non-immunogenic. [26]

▲ TYPES OF NANOPARTICLES

valent liposome nm. [25]. These

2.1.4. Compact Polymeric Nanoparticles :-

Compact polymeric nanoparticles are nanostructures produced exclusively of innate or artificial polymers. They are generally more stable than liposomes favouring continuous localised drug delivery for weeks, with restricted drug leakage. [23]. In these polymeric nanostructures, the therapeutic agent can be eventually linked covalently. Alternatively, it can be adsorbed at the NPs surface or dissolved or enmeshed within the nanoparticles structure (nanospheres) or encoated inside a polymeric shell (nanocapsule). [23, 27]

2.1.5. Hybrid :-

An transitional type of NPs is the core-shell polymer-lipid hybrid NPs. In its structure a biodegradable hydrophobic polymeric core and a lipidic outer monolayer are present [28]. Alternatively, an inner polymeric core encompassed by an external lipid bilayer can be utilised [29]. Core-shell polymer-lipid hybrid NPs bring about the complementary

characteristics of both complexes, namely higher stabilised, enriched drug coated yield and superior in vivo cellular delivery efficacy [30].

2.2 Inorganic Nanoparticles :

2.2.1. Fullerenes :-

NPs can also be easily made of carbon molecules with various highly symmetric and stable forms, called fullerenes (allotrope of carbon)[31]. Buckminsterfullerene (C₆₀), the most referred to as fullerene, is a rigid icosahedron with 60 carbon atoms. In its structure, single bounds form pentagons and double bounds form hexagons[32]. Fullerenes disadvantages, such as the low solubility in organic solvents, are overcome by their unique optic, electric and magnetic properties (such as superconductivity[33]), rendering them important devices in medical diagnosis and imaging [33].

2.2.2. Inorganic materials, for example **gold, silver, platinum and silica**, can also be used to produce NPs.

2.2.3. Quantum Dots:-

Recurrently, quantum dots are delineated as “artificial atoms”. These were one of the first nanotechnologies to be extensively used in biological sciences and are expected to treasure some applications in the near future, in a number of commercial consumer and clinical products (Klimov 2007, Valizadel et al.2012).[34,35] QDs present exclusive luminescence properties and electronic characteristics, such as broad and uninterrupted absorption spectra, thin emission spectra and high light stability (Bruchez et al 1998).[36]QDs manifest high wherewithal for identifying cells, discover particles, and accumulating solar energy. However, before we look into the advantage of this new particle we should first figure out the risk of its toxicity to organisms.Nair et al (2011) observed that when rice seeds were treated with CdSe QDs, it was detected that the germination of the seeds were constrained [37]. According to Hoshino et al 2004, QDs can lead to DNA damage and decelerate the germination of cells in culture.[38] It was also noted that cell damage and cell death can be induced by mercapto-undecanoic acid QDs (Shiohara et al 2004)[39].

Although the vulnerability of single walled carbon nanotubes to plant induced benign effects, the cluster of QDs to the nanotubes effectively transform the viability of the tomato plants by significantly stimulating leaf senescence and inhibiting root formation (Alimohammadi et al 2011).[40]Furthermore, it was noted that, when *Arabidopsis* is unveiled to QDs, it induce oxidative stress, as conceded by changes in the GSH/GSSG ratio (Navarro et al 2012).[41]

3.Characterization of Nanomaterials

Characterization of nanomaterials being a part of nanometrology, handles the characterization of the physical and chemical properties the nanomaterials. This helps us to

evaluate nanomaterial in their degree of toxicity and usefulness in different genres of life. However, there are several microscopic and spectroscopic mechanisms into practice. Metrics, taken into consideration for the characterization of different nanoparticles are size and dispersion, shape, chemical composition and crystal structure, surface area, surface chemistry and charge, solubility.[42]

3.1. UV spectroscopy - It is the method by which light is incorporated by the sample and scattered in the sample is quantified. The measurements of spectroscopy are compared at different wavelengths. Nanoparticles having optical properties can be easily characterized by this method. The measured spectrum is compared expected spectrum with the help of many numerical tools.

3.2. Transmission Electron Microscope (TEM) - It is a tool in microscopy to produce highly magnified images produced by transmission of electrons in the sample. Electrons instead of light is used to illuminate the images produced. Background of the sample plays an integral part in the characterisation of nanoparticles via TEM. Principle behind this is the high electron of the nanoparticles that make the imaging feasible. It is most used to measure metal based nanoparticles and few carbon based nanoparticles like carbon nanotubes, quantum dots, magnetic nanoparticles etc.

3.3. Dynamic Light Scattering (DLS) - This is an important characterisation of nanoparticles based on their colloid nature. Light gets scattered in the colloidal solution and by the analysis of the brownian motion of the colloidal particles. The hydrodynamic diameter measured in this case is larger than the size analyzed in TEM.

3.4. Zeta Potential - It is a physical characterisation that help to quantify the total surface charge of the nanoparticles. It is a measure of the discrepancy in potential between the bulk fluid in which a particle is immersed and the layer of fluid covering the oppositely charged ions on the nanoparticle surface.

3.5. Atomic Absorption Spectroscopy (AAS)- It is the analytical technique that helps in determination of mass concentration of the nanoparticles in consideration. It works on the principle of atomic absorption from light to a specific wavelength. The amount of energy hence absorbed is related to the number of atoms on the light path.

3.6. Fourier Transform Infrared [FTIR] spectroscopy – Determination of the nature and structure of functional group can be conferred by measure of infrared wavelength against light wavelength. This spectra scrutiny can highlight the optical properties of nanoparticles.[43]

3.7. Energy Dispersive X ray analysis (EDX) – it is an analysis technique for surface inspection on nanoparticle, from all sides and proportions. Thus, giving an overall mapping of the nanoparticle surface. It is laborious technique because of the low penetrating power of the X rays.[44]

3.8. Atomic Force Microscope (AFM) – It is a non destructive analyzing technique, with a very high spatial resolution. This makes it ideal for nano range analysis. Images are

generated by the feedback system found between the optical detection and piezoelectric scanners.[44]

3.9. Scanning Tunnelling microscopy (STM) – this helps us to study particle surface from scale lateral resolution. It works on the principle of quantum tunnelling, which implies that on the contact of conducting tip to the surface, electrons are tunnelled through the vacuum between them.[44] It is widely used in the characterization of carbon based nanoparticles.

4. Mechanism and Principles in Nanotechnology

Consumption, translocation, and assemblage of NPs dependent on the plant system and the size, kinds, chemical conformation and stability of the NPs. The consumption, bio metabolism and transfer of various NPs in a plant architecture has been earlier demonstrated [45]. The intake, assimilation, and the transfer of organic matter, suspended fullerene C₇₀ and MWCNT in rice plants were examined in the past [46]. Black agglomerates of C₇₀ were observed. These stocks were obtained more in the seeds and roots as scrutinised to the stems and leaves of the rice. It has been reckoned that the presence of aggregates of NOM-C70 in leaves led to the transference route of water and the nutrients through the xylem tissue. In the cell membrane specific ion transmitters have been concluded for NPs ascended by the plants [47]. Because of the low surface friction of CNTs, the passage of organic substances into the cytoplasm can be given out [48]. Interplay of ryegrass with NPs has been recorded. The scanning electron microscopy studies confirmed the adsorption and aggregation of the NPs on the root surface [49].

TEM images of root cross-sections of the ryegrass also displayed the occurrence of particles in the apoplast, cytoplasm and nuclei of the endodermal cells. Birbaum [50] detected CeO₂ NPs employed on corn leaves, which were absorbed by the leaves, but not transferred to new leaves. Zhu [51] informed the uptake of Fe₃O₄ NPs by pumpkin seedlings in husbandry culture using a vibrating sample magnetometer. NPs were detected in roots, stems, and leaves of the plants. It was compelling that no uptake was noted when plants were germinated in soils and a reduction in uptake was noticed, when grown on sand. In accordance with the Fe₃O₄ NPs, growth medium was not taken into prominence, in the soil and sand grains. Incapability of uptake of lima bean plant species, was noticed on treatment with Fe₃O₄ nanoparticles. the ascent and transmission of Cu NPs in mungbean (*Phaseolus radiata*) and wheat (*Triticum aestivum*) were examined by Lee *et al.*, [52] in the agar growth medium.

It was inferred that the Cu NPs could navigate through the cell membrane and aggregated in the cells. *Cucurbita pepo* when administered with Ag NPs, the Ag concentration in the plant shoots was found to be 4.7 times higher in the plants than those treated with large amount of Ag powder [53]. Cases of biomagnification has been reported in algae and tobacco via nanoparticle treatment[54,55]. NPs assimilated in the cells may be transferred by means of apoplast or symplast through plasmodesmata. However, the exact methodology by which plants take up NPs and plant specific accumulation of NPs is are still undiscovered and remain unexplored.

Nanomaterial unveils in plant system through numerous pathways, such as peripheral emission from manufacturing orifice in sewage treatment plants [56,57]. In nutrient rich organic compounds from sewage water treatment domain, pesticides employed to agronomy, dyes, textiles, private health purpose, and sudden descent of materials at the time of manufacturing consumer products[58]. When nanomaterials certain excretion from

diesel release, sewage water appear at agricultural land have an increased capability to pollute soil, fertility transfer into surface/ground water, and interact with normal biota. Further these nanoparticles can also be commuted to an aquatic system by rainwater bodies miscellany may lead to its buoyancy's imperceptible leaching.

4.1. Role Of Engineered Nanomaterials In Plant Growth By Various Ways –

4.1.1. Carbon Based Engineered Nanomaterials :

An increased production of carbon-based nanomaterials has paved way to its potential discharge in living systems, either willingly, accident in content, and showing more potential of the unsatisfying environmental effects [59]. Amid the carbon nanomaterials, the most volitional materials are fullerene C_{70} , fullerol ($C_{60}(OH)_{20}$), and carbon nanotubes. Generally carbon-based nanomaterials are measured highly hydrophobic with the proclivity to aggregate, and expected to settle in the living system [60].

4.1.2.Fullerene on Plant Growth :

The fullerene as an aggregate of black blend is more profuse in seeds and roots growth as compared to the leaves and stems of rice seeds [61]. While in mature plants, favourable transportation from the roots to aerial part of plant has become more prominent. Thus, fullerene agglomerates mainly occurring inside or close to the the stems vascular system and leaves, whereby the roots have been deprived of fullerene [62]. The conjunction of fullerene in leaves also states that pathway of nutrients and water through the xylem [63]. It is heralded that individual fullerene nanoparticles invading the root surfaces in accordance to osmotic pressure, capillary forces, and cellular pores through the intercellular plasmodesmata, or by using very much synchronized symplastic routes [64,65]. The fullerene moieties with a calibre of a smaller capacity than the pore size of the cell wall could easily flow and reach to the plasma membrane.

4.1.3.Multi Walled Carbon Nanotubes (MWCNTs) On Plant Growth :

The Multi walled carbon nanotubes (MWCNTs) are 1mm long and 20 nm in diameter [66-69]. And using the seeds and roots arrangement through the genesis of new outlets and water intake in order to build tomato seedlings [70,71]. The rhizome was envisioned before finally racing the epidermal and root hair cell walls and cap of the seedlings [72]. It is also heralded that MWCNTs are impregnated in tomato seeds and enrich the emergence rate by enhancing the seed water intake. The MWCNTs upraised the maturation of seed to up to 90% in 20 days juxtaposed to 71% in the standard specimen and the vegetational biomass [73].

4.1.4. Single Walled Carbon Nanotube (SWCNTs) - On Plant Growth

The acreage of single walled carbon nanotube (SWCNTs) is about 1 to 2 nm in diameter and $0.1 \mu\text{m}$ in length. water column stoutness in cucumber seedling after handling for 84 h the SWCNTs were found cohere to the outer surface of the main and secondary roots

[74,75]. However, coexisting information are lacking to confirm the translocation of SWCNTs from the root systems to the aerial parts of the plant [76].

4.1.5. Titanium oxide (TiO₂): On Plant Growth

The titanium oxide Nanoparticles (TiO₂) are substantially deployed in daily life products, but the probing of their ingestion and translocation in the plant is limited, particularly on food crops [77-79]. TiO₂ size (<5nm), TiO₂, tend to form a chemical bond with most of the non-conjugate ordinary organic matter, translocate, and following the tissue and cells' [78-80]. TiO₂ nanoparticles with nitrate unsheltered in Soybean (*Glycine max*), intensify the ability to utilise, and stimulate the inhibitor system. For instance, TiO₂ nanoparticles corollary plants showcased 73% increased dry weight, three times raised photosynthetic rates, and 45% chlorophyll revampment as compared to the domination over the cultivation phase of 30 days.

Some studies professed that the TiO₂ nanoparticles low high prospective in sponge up of inorganic nutrients, escalated the disintegration of organic substances, curbing by oxygen free radicals formed during the photosynthetic process, thereby manipulating the photosynthetic rate [81,82]. It is divulged that toxicity of TiO₂ nano particles low due to particle collection and ensuing accumulation. The toxic effect of TiO₂ nano particles is feasibly not imputed by the released Ti²⁺ ions from particles that are capriciously manifested by the finite disbandment of Ti from a TiO₂ specimen [83]. Genomic DNA ascertainment was perceived in the root tips of cucumber after seven days and designated that plants served with 2000-4000mgL⁻¹ of TiO₂ nano particles reduced the genomic DNA compare to the control sample [84,85].

4.2. Mechanism of intrusion of Nanoparticles

Utilizaion of nanoparticles for the designated production of substances hasattained special concentration and is of major insight in the treatment of plant diseases. Examinations were done for observing both the perforation and the movement of iron-carbon nanoparticles in plant cells have been examined in living plants of *Cucurbita pepo*. The nanoparticles were surveyed *in planta* using two different application methods, injection and spraying, and magnets were used to preserve the particles in movement in specific areasof the plant. The sole experimental set up, using correlative light and electron microscopyprovided evidence of intracellular insertion of nanoparticles and their displacement from the application point. Long range movement of the particles through the plant body was also diagnosed, particles having been found near the magnets used to deactivate and centralise them.

Furthermore, cell reaction to the nanoparticles occurence was dignosed. Nanoparticles were competent of puncturing living plant tissues and itinerating to different regions of the plant, although movements over short distances seemed to be advocated.

The outcomes show that the use of carbon coated magnetic particles for administered delivery of substances into plant cells is a attainable application.[86]

5. Nanotechnology and Plant Growth

5.1. Phytotoxicity

Agriculture and plant culture is an prominent ae of development, which has a continuous ardent need of improvement. The growing prominence of the efficiency of nanoparticles has paved it way to be helpful in the progress of agrarian fields[87]. Nanoparticle usage strives for a betterment in application techniques and considerable reduction in amount of plant protection products(PPP), also to mitigate the dearth of nutrients evident during fertilization and other plant processes[88,89]. The carbon based nanoparticles have shown a considerable amount of notability in plant nanotechnology.among them,Multi-walled carbon nanotubes (MWNTs) are the nanomaterials that have grown to be remarkable interest in research field and technological development due to their unique caricature and peculiar properties. However, repetitive usage of nanomaterials in plant architecture can culminate into bioaccumulation with may ultimately lead to toxicity. This gives rise to the concept of phytotoxicity of nanomaterials.[90] Nano-encapsulated agrochemicals are delineated in such a way that id encompasses all the required properties and necessary effective concentration with a judged evaluation of time controlled release in plant body with escalated target activity and little ecotoxicity with safe and elementary mode of delivery for avoiding repeated application[91-93].

The prime factors guiding the effects of NPs on plants are the properties of the nanomaterials like concentration, size, category, stability [94] Some studies have shown that phytotoxicity aggravates with particle size. For example, Yasur and Rani (2013) affirmed that all Ag NP treatment groups had no significant effect on the growth of castor, but in the treatments where the Ag was emergent in its bulk form, subdue in the process was observed. This was affirmed by the work of Lee et al. (2010).

Phytotoxicity is a notable cognition in comprehending the potentialhabitual impression of nanoparticles. Profuse analytical data have shown thatmulti-walled carbon nanotubes (MWNTs) are pernicious to plants, but the potential impacts of revelation remain perplexed. The intention of the current survey was to gauge estimatedphytotoxicity of MWNTs at different specific levels with red spinach, lettuce,rice, cucumber, chili, lady's finger, and soybean, based on root and shoot enhancement, celllysis, and electrolyte crevice at the seedling stage. 15 days later of agronomic culturing,the root and shoot extents of red spinach, lettuce, and cucumber were considerably diminished leading to exposure towards particular MWNTs. Indistinguishable toxic sequels occurred regarding cell death and electrolyte leakage. Red spinach and lettuce greatly was responsive to MWNTs, followed by rice and cucumber. Nominal or no adverse effects were observed for chili, lady's finger, and soybean.[95]

5.2. Bacterial effect

With the advancement of Nanotechnology, it is necessary for us to pave non toxic and eco friendly ways of the bio synthesis of nanoparticles. Metal and engineered based nanoparticles, although widely used, face the criticism of being toxic after a particular or excessive usage. However, techniques lilke green synthesis of nanomaterials, i.e. nanomaterials synthesized from plants, which are the green sources. This field of nanotechnology accelerated a new radar of research in the botanical aspect in this feild.

This however, was then further redirected to support the school of thought that nanomaterials can be synthesized via living or biologically sound organism.

This gave rise to research regarding synthesis of nanoparticles via micro-organisms. Microbial synthesis of nanoparticles is extremely advantageous and takes place at an ambient environment and temperature. The size and shape of the nanoparticles can be maneuvered with respect to the micro organism used. Silver nanoparticle is usually synthesized, owing to its remarkable anti bacterial property.[96] These nanoparticles, if administered in plants will ensure, bacterial protection and prevent infection. Also, Zinc oxide nanoparticles are beneficial [97,98].

These nanoparticles are anti pathogenic and hence can be used in a eco friendly scenario. Soil microbes are an integral part of the ecosystem and contribute hugely to the ecological design[89]. Synthesis of Ag ,Zn NPs are usually done from pure cultures of *Escherichia coli*, *Staphylococcus*, *Bacillus*, and *Pseudomonas*,etc. in laboratory situations[100-104]. Incoherence of growth in soil bacterial communities have been reported.[104-106] concentration of nanoparticles in restricted amount can result in being sub lethal.[107]

Effectivity of nanoparticles is attributed by have high surface volume ratio, making it highly metabolically effect.(Ying,2011) . The reaction of nanoparticles with the bacterial colony can be assessed for enlightening the extent of scope of nanotechnology related to biological synthesis and its farfetched effects.[98] A significant differences were not observed when bacteria like *Burkholderia kururiensis* was growth evaluated in the presence of NPs. On the other hand, *Pseudomonas fluorescens* showcased a slower exponential growth phase when exposed to nanosilver leading to bioaccumulation of these nano subunits[108]. However, a sublime effect of NPs on bacterial development and plant bacterial interactions has been reported in frequent cases.

5.3. Stress

Many chemical and physical changes are endorsed in the plant system, by the interaction of nanoparticles and plant tissue system. Nanoparticles ranging in the size from 1-100 nm can of varied size and have corresponding effects. Smaller the nanoparticles, greater being the surface, can have large scale impact on the plant tissue. However, larger nanoparticles (NP) reach upto the restricted area of the plant tissue system, agitated by the cell thickening and hence have a controlled effect on the plant body. Thus, we can say the smaller NP which are majorly responsible for physical and chemical changes bring about a change in stress impact of the plants.

Metals have a integral role in the life process of the plant working system. Metal derived nanoparticles have showed great prominence in altering plant process and internal environment. This is due to the fact that metals have immense significance role in the physiology in the plant architecture. Chemical interplay results in the production of reactive oxygen species (ROS), disturbance of ion cell membrane transport activity , oxidative deterioration , and lipid peroxidation.[109-112] By different metabolic processes in the plant

body, the produced oxygen is reduced to different reactive species like peroxide, superoxide etc. The continual removal mechanism of ROS can get deterred from different environmental and chemical changes. Thus, administration of metal nanoparticles has revolutionized the combatting of biotic and abiotic stresses of the plant body. Infringement of nanoparticles in plant body can increase stress conditions. Both metal and carbon based nanoparticles are able to deliver ROS induced stress. However, few metallic oxide stand as exception.[116-117] Also, the engineered nanomaterials (ENM) pose many adverse effects on the plant growth and may also culminate into nutrient depletion. These may in result to the phenomena of nanoparticle stress conditions.[114-115] A check on photosynthetic efficiency can highlight the stress conditions of the plant body. Nanoparticle stress can occur on physiological and chemical aspects. Some, carbon nanomaterials effect the plant on genetic and proteomic levels. Out of the metal based nanoparticles ,AgNP deserve special mention as its exposure can bring about signaling and metabolic stress in the plants.[113]

Abiotic stress can be induced by CuO and AgO nanoparticles, in greater quantity as accumulation of metals can increase toxicity. However, in little amount it can result in enhanced plant growth in different spheres[117-119]. Bacterial biofilm formation in the root process is not perturbed by the inception of metal based nanoparticles, in spite of the chance of toxicity which can be caused by the presence of metal ions.[122] Drought and excessive salinity have posed a great threat to the agricultural land productivity. Hence, Na(sodium) being a very important metal in the soil system, can help in the improvement of the deteriorating scenario via the use of nano-reclaimants like nano gypsum, nano calcium and magnesium compounds, etc.[123]

5.4. Influence of Metal Nanoparticles :

Nanoparticles of gold (Au), silver (Ag), copper (Cu), zinc (Zn), aluminium (Al), silica (Si), zinc oxide (ZnO), cesium oxide (CeO₂), titanium dioxide (TiO₂) and magnetised iron (Fe) have found its usefulness in agro-economic areas.(Josef and Katarina, 2015) [124].

For the increment of food production, it is imperative to use various technologies in agriculture. Fertilizers composed/manufactured with nanotechnology have extraordinary significance. This study was endeavored to determine the impact of nanotechnology liquid fertilizers on plant growth and development. (Melek et al., 2014) [125]

5.4.1. Copper Nanoparticles :-

CuNPs can be synthesized from onion (*Allium cepa*) extracted and ultrasonified at 100 degrees Celsius in a water bath. The translucent yellowish green colour indicated the transformation of copper ions into CuNPs. It was reported that wheat crops when administered with CuNPs in diverse concentrations (15-55ppm), 35ppm gives better results in terms of parameters like chlorophyll content, shoot length, root length, germination percentage, dry and fresh root weight, in comparison with the control. Beyond 35 ppm, it was noted that the growth of wheat crops decreased. Hence, it can be deciphered that

CuNPs have the capability to augment the growth of wheat crops till a certain concentration. [126]

Copper –carbon core shell nanoparticles increase Cu intake in 2 year old bald cypress (*Taxodium distichum*) seedlings but there was no considerable effects on the growth specifications of the seedlings after 25 weeks of NP administration (Qi et al). Qi et al also showed that wood administered with copper-carbon core shell nanoparticles are highly defiant to blue stain (*Ophiostoma minus*) and white rot (*Trametes versicolor*) fungi; and Formosa subterranean termite (*Coptotermes formosanus*). [127]. Contrarily, Lee et al observed toxicity of CuNPs in two crops species namely, mung beans (*Phaseolus radiata*) and wheat (*Triticum aestivum*), as manifested by declined seedling growth rate. [128]

5.4.2. Silver Nanoparticles :-

Silver NPs have exclusive characteristics such as high surface areas, catalytic ability, surface energy, ample reactive sites and stable adsorption, can have vital effects on many organisms chiefly plants that fundamental sources of all ecosystem [129,130]. The catalytic action has been verified by expanded degeneration of starch and amylase enzyme [131]. AgNPs also demonstrate photocatalytic actions. [132]

Withania somnifera, a significant medicinal plant could not comply the demands of classical medicine practice, because of seed dormancy. The seed dormancy was subdued by administering the *W.somnifera* seeds with NaDC-capped AgNPs for 90 mins, produced germination that was remarkably higher than the control. [133].

AgNPs has been indentified to hinder the ethylene activity.[134]. Ag eradicates undesirable microorganism in farmers soil and hydroponics system. It is applied as foliar spray to forbid fungi, rot, moulds and various other plant diseases. Furthermore, Ag is an excellent plant growth enhancer , including silver salt, silicate and water soluble polymer to radioactive rays.[135]

The administration of AgNPs in the fenugreek seeds enhanced seed germination, mean germination time, seed germination index, seed vigour index, seedling fresh and dry weight.[136].

5.4.3. Titanium Dioxide Nanoparticles :-

TiO₂NPs has a great significance on bacteria, algae, plankton, fish, mice and rats but research should concentrate on the effects of TiO₂NPs on plants which is still lacking.

TiO₂NPs augment seed proliferation and endorse radical and plumule growth of canola seedlings (Mahmoodzadeh et al 2013) [137]. As reported by Jaberzadeh et al 2013, TiO₂NPs amplify wheat plant growth. [138]

TiO₂NPs coordinate enzyme actions concerned in nitrogen metabolism such as nitrate reductase, glutamate dehydrogenase, glutamine synthetase, and glutamic pyruvic transaminase that support the plants to absorb nitrate and also aids the transformation of

inorganic nitrogen to organic nitrogen, materialize in the form of protein and chlorophyll, that escalate the fresh and dry weight of plants (Mishra et al 2014, Yang et al 2006) [139,140]

TiO₂NPs enact as photocatalyst and promote oxidation-reduction reaction. (Crabtree 1998) [141]. TiO₂NPs induced photosynthesis and nitrogen metabolism and hence immensely augment the growth of spinach at a concentration as low as 20mg/l. [140,142,143]

5.4.4. Gold Nanoparticles :-

Researchers noted that AuNPs enhance toxicity in plants by restraining aquaporin function, a group of proteins that support the ascent of a large range of molecules counting water molecules also. (Shah and Belozeroval 2009) [144]. Nevertheless, researchers like Barrena et al (2009) in lettuce and cucumber [145],Arora et al (2012) in Brassica juncea [146], Savithamma et al (2012) in Boswellia ovalifoliolata [147] and Gopinath et al (2014) in Gloriosa superba [148] demonstrates that AuNPs boost seed germination.

Neomycin phosphotransferase II gene was imported into soyabean genome by means of DNA- coated gold particles. (Christou et al 1998). [149] The benign effects of AuNPs require further analysis to delve into the physical and molecular methodologies. According to Kumar et al (2013), AuNPs have a paramount role in seed germination and antioxidant scheme in Arabidopsis thaliana and transformed levels of microRNAs expression that governs various morphological, physiological and metabolic processes in plants. [150].

5.4.5. Sodium Silicate And Silica Nanoparticles :-

Silica is another most predominantly found element in soil which has benign effect on living and non living, and escalate stress resistance in plants. It can boost the production and product quality, abate evaporation of perspiration, raise the stimulation of a few anti-oxidant enzymes and downturn the receptivity of some fungus. Succeeding oxygen, silica is an effective element in earth, that is non-mobile in plants. Despite the fact that silica is not paramount for plants, higher plants require it for having optimum growth (Richmond & Sussman 2003, Ma et al. 2004, Currie & Perry 2007).[151] The influence of silica on plant is akin to intransigence against biotic and abiotic stress (Ma & Yamiji 2006, Liang et al. 2007).[152] Silica is a crucial constituent against salt stress (Zhu et al.2003) [153], manganese toxicity (Shi et al. 2005) [154], boron toxicity (Gunes et al. 2007)[155] and cadmium toxicity (Shi et al.2010) [156], by means of altering the action of antioxidant enzymes.[157]

5.4.6. Zinc Oxide Nanoparticles:-

Zinc has been viewed as crucial micronutrient for metabolic action in plants. It balances the various enzymatic action and enforced in biochemical reactions leading to the development of chlorophyll and carbohydrates. ZnNP is used in discrete agricultural experiments to figure out its outcome on growth, germination and other distinct attributes.[158]

Information proposes that zinc oxide nanoparticles fortify plant growth and development. It has been noted that in peanut, soyabean, wheat, onion, lessened concentration of ZnONP shows benign effect on seed proliferation.[159-162] It has been tested that diverse

combination of ZnONPs on cucumber, alfalfa and tomato; showed incremented in seed germination only in cucumber.[163]

ZnONPs are likely pertinent in the field of bio-imaging and in the construction of printable electronic devices. These NPs can be allowed to be a choice for the creation of antimicrobial textiles. They are also utilized in nano-medicine for oncological purposes.[164]

6. Implementation of Nanoparticle to Enhance Plant Growth and Yield

6.1. Nanoparticles As Growth Promoter

In the previous years, the experimentalists have mentioned the consequences of NMs on cultivation and development with the aim to publicise their utility of agricultural applications. Reactivity of NPs with plants caused various corporeal and physiological changes, based on the attributes of NPs. Efficaciousness of NPs depends on their dissolution and it varies significantly. Coherency of NPs is determined by their chemical composition, size, surface area, reactivity, and the concentration at which they respond positively [165]. NPs pose both constructive and dismissive influence on flora. However, it is the objective to establish the positive influence of NPs on seed harvesting, photosynthesis and growth. NPs commonly confronted in the agricultural field fall into respective groups: carbon NMs, metal NPs and metal oxides NPs.

6.1.1. Effect of carbon nanomaterials on plants

Among the NPs, carbon NMs have been acknowledged with a significant place due to their individualistic mechanical, electrical, thermal and chemical properties. The tomato seeds when came in contact with MWCNTs, seed harvest and plant biomass incremented significantly [165]. It has been hypothesized that penetration of the cell wall by NPs leads to the rise in water uptake by seeds which enriched germination percentage. Srinivasan and Sarawathi [166] supported that the single walled-CNTs (SWCNTs) act as nanotransporters for portage of coloured molecules and genetic materials into plant cells. Studies state MWCNTs enriched efficiency of water uptake along with Ca and Fe components uptake incremented the germination and growth [167,168]. Seed cultivation improved in barley, soybean and corn when treated with MWCNTs, examiners using Raman spectroscopy and TEM detected nanotube aggregates inside the seed coat [169]. Oxidized MWCNTs raised cell elongation in rhizome and favoured dehydrogenase action [170]. Impact of water soluble carbon dots which induces certain modifications such as seed germination, root growth and elongation. However single walled CNT can cause Phytotoxicity and related destructive attributes.[171-173]

6.1.2. Effect of metal nanoparticles on plants

It's observed that metal NPs enhanced plant development. AgNPs impacted the rhizome size in maize and cabbage plants in contrast with AgNO₃ [174]. Au NPs affected the leaves content, area, altitude and glucose and chlorophyll levels that gave improved crop yield [175,176]. AuNPs play an integral role to play on germination and antioxidant levels in *Arabidopsis thaliana* [177]. Au NPs enhanced seed growth in *Boswellia ovalifoliata* [178]. Biologically calibrated Ag NPs induced synthesis of protein and carbohydrate and degraded the total phenol content in *Baopa monnieri* [179]. Root length increased in barley exposed to AgNPs [180]. Spinach seeds seeped in Ti NPs solution raised fresh and dry weight driving

the contents of overall chlorophyll and protein in leaves [181]. Increasing the two fold caused rise in height and fresh weight of duckweed when reacted with Ti NPs at 0.5 gL⁻¹ conc. [182].

6.1.3. Effect of metal oxide nanoparticles on plants

Numerous studies on the metal oxide NPs' effectiveness on plants are chronicled. Nanosized TiO₂ favoured plant growth when seeds were allowed absorb in NPs or sprinkled with NPs [183]. Parsley seeds treated with nano anatase, enriched cultivation, root and shoot length and chlorophyll levels of the seedling [184]. The maturation rate of salvia enhanced on seeds exposure to TiO₂ NPs [185]. TiO₂ and SiO₂ NPs blend elevated the nitrate reductase activity and favoured the antioxidant system in soybean [186]. In spinach, chlorophyll production, photosynthesis and plant dry weight intensified when treated with TiO₂ NPs [187,188]. Root elongation favoured at certain specificity of ZnO NPs in soybean [189]. Iron oxide NPs enhanced root elongation in pumpkin [190]. Analysis showed iron oxide NPs incremented soybean pod and leaf dry weight [191]. ZnO NPs led to plant germination in the peanut plant [192]. Suriyaprabha [193] described that seed germination improved using nano SiO₂ which led to advanced nutrient flow to maize seeds.

6.2. Nanoparticles In Disease Suppression :

Viruses, bacteria, fungi and nematodes are mainly liable for plant diseases leading to decreased yield and abysmal quality of plant products. Various propositions to manage crop disease are being used including genetic breeding, cultural schemes with sanitation, host indexing, enhanced extermination protocols, newer pesticides, and integrated pest control. Studies state that NPs were applied to extirpate pathogens which incremented crop growth. Jo et al [194] concurred that Ag NPs in 200 mg/l conc. reduced 50% colonies of pathogenic fungi that caused virulence in ryegrass. Lamsal et al [195] stated that use of Ag NPs favoured the disease eradication. Combination of Ag NP with the fungicide fluconazole were figured to be consequent against *Candida albicans*, followed by *Phoma glomerata* and *Trichoderma* sps [196]. ZnO NPs diminished growth significantly of *Fusarium graminearum* in mungbean broth agar [197].

MgO NPs evinced considerable antimicrobial action due to powerful interplay with a destructive layer of bacterial membranes [198]. Chemically obtained Cu NPs denoted increased infectious fungal impedance in contrast to the fungicide bavistin [199]. Jo et al. [200] stated effective potencies for obstruction of colony production by silver compounds on *Bipolaris sorokiniana* was enhanced than that on *Magnaporthe grisea*. Silver compounds impeded colony formation of *B. sorokiniana* by 50% at optimum levels. Considerable decline in mycelial growth was recorded from spores nurtured with silver NPs [201]. Silver NPs largely declined the number of cultivational fragments relatable to the control at 24 hour germination of spores with a 2.5 ppm solution of NPs. Field tests were conducted with silver NPs (WA-CV-WA138) at various dissolutions to diagnose antifungal activity. The maximum obstructive rate for the growth of pathogen on cucumber and pumpkins pregnated with 100 ppm silver NPs [202]. Gopinath et al [203] in his scrutiny described antimicrobial and larvicidal activity of biologically intermixed Ceo₂ NPs (Fig. 3).

6.3. Nanopesticide And Nanoherbicide :


Pesticides and herbicides are generally used in agriculture to get better crop yield and efficiency. Currently the destructive aspects of conventional pesticides and herbicides on environment are under argument. The bane of pesticides are favouring the growth of pathogen and pest resilience, degrade nitrogen fixation, declination in soil biodiversity, contributes to bioaccumulation of pesticides, pollinator decline and destroys habitat for birds [204]. Hence, utility of NPs reconciles these problems to most extent, its application with herbicides diminishes the amount of herbicides required for weed expulsion. Having the dynamic and necessary delivery system, herbicides are sprayed in the soil in accordance with the soil condition (Gruere *et al*) [205]. Ag NPs have pesticidal affair against infectious fungi, declared to have obstructive consequences on conidial harvesting of genus *Raffaelea* which leads to the imparment of oak trees [206].

6.4. Nanofertilizers :

Nanofertilizers are referred to as NMs which produce nutrients to the plants or provide assistance to accelerate the proceedings of traditional fertilizers. Replacing nanofertilizers with standard fertilizer is befitting as its exercise is to vent out nutrients into the soil stability and in a restricted manner, thereby averting water contamination [207,208]. Hydroxyapatite ($\text{Ca}_5(\text{PO}_4)_3\text{OH}$) NPs of 16 nm in dimension, integrated by Liu and Lal [209], evince fecundity effect on soybean. Many survey suggest the application of nanofertilizer reflect upon the constructive effect concerning better crop production along with environmental hazard. The application of NPs incremented the growth rate and seed harvest by 33% and 20% respectively, contrary to typical P fertilizer. The result discerns that the underground system of soybean can hydroxyapatite NPs as an influential P nutrient source. Soil amended with metallic Cu NPs considerably has an accrual effect on 15 day lettuce seedling exhibiting a growth of 40% and 91% respectively [210].

Most surveys throws light on the properties of NPs leading to the revelation of the ability of NPs which can enter the plant cells and transfer DNA and chemicals inside the cell [211-213] surveys gave a podium for estimating that NPs can also provide nutrients to the plants in the form of fertilizers. The nano-organic iron chelated fertilizers showcases high absorption, rise in photosynthesis and elongation of laminal surface area [214]. Additionally, nanofertilizers have immense effect on the soil as nanofertilizers can decrement the harshness of the soil and degrade the recurrence of fertilizer application [215]. Manikandan et al stated that nanoporous zeolite applied on N fertilizer might be used as subordinate structures to enhance the potency of N in the crop production system [216]. . Nanofertilizers due to their properties driven features have immense impact in sustenance of agriculture [217]. Therefore fr

of nanofertilizer re
minimise the soil to

 **EFFICACY OF NAOFERTILIZERS & NANOPESTICIDES**

nents,
cy.

6.5. Biosensors :

NPs can be employed as a detective medium for diagnosing of plant pathogens. This analysis is in nascent stage in agriculture. NPs sometime deployed as a examinational tool to identify compounds which could help indicate disease. Fundamentally biosensor has emerged from the grouping of a ligand-receptor binding reaction to a signal transducer. It comprise a inquest, bioreceptor and transducer. The interplay between analyte and bioreceptor is structured to perform an effect estimated by transducer, metamorphose the

documentation into electrical signal (Fig. 4). Lopez et al [218] recorded that nano-chips are known for identifying single nucleotide alterations of bacteria and viruses. These nano-chips involve fluorescent oligo apprehension probe through which conglomeration can be figured out. *Xanthomonas axonopodis* leads to bacterial spot disease in Solanaceae plant. Yao et al [209] utilised fluorescence silica NPs in amalgamation with antibody to diagnose the microbes displaying potential of NPs for causing disease. It is now evident that nanobiosensors can be used bounteously for intuitive diverse array of fertilizers, herbicides, pesticides, insecticides, pathogens and thus can favour sustainable agriculture by enriching crop productivity.

6.6. Distribution of Nanoscience in different fields of application.

Miscellaneous use of pesticides and fertilizers leads to environmental adulteration, transpiration of pest, disease, virulence and biodiversity loss. Nanotechnology, through the nanomaterial based characteristics, has potential agro-biotechnological applications to propitiate these problems.

The documentation concerning to the impact of nanotechnology in plant and soil system illustrates that, Nanomaterials may help in the formulations of nanomaterials-based pesticides and insecticides, elevation of agricultural yield using bio-conjugated nanoparticles (en coating) for nominal emission of nutrients and water, nanoparticle-mediated gene or DNA transfer in plants for the evolution of insect pest-resistant miscellany and application of nanomaterials for preparation of different kind of biosensors, use of nano clay composite superabsorbent for higher retentivity of soil moisture and essential plant nutrient is effective in revaluation of the drought stress tolerance in the crop.

The nanoclay composite with unprecedented easy outflow and water-retention capability, nontoxic to soil and environment, could be specially utilitarian towards agricultural and horticultural applications for high input use proficiency.

7. Advancing Nanotechnology in India

The endeavour to urge the emergence of Nanotechnology in India commenced years ago. Nanotechnology has always gained enough momentum when it is reckoned to constitute a beneficial effect on India. Numerous initiatives have been undertaken in order to make Nanotechnology more prosperous in India. The impact of Nanotechnology is varied in different realms of life. Instances to cite, on how Nanotechnology has been evolutionized in India.

Dating back to 1998, when the 9th Five Year Plan (1998-2002) was established that made its maiden declaration that national facilities and core groups were build to stimulate research and studies in frontier areas of science and technology which comprised superconductivity, robotics, neurosciences and carbon and nano materials. Planning Commission confirmed the quantity of such research and developmental programmes under standard research (GOI 1998).

Nevertheless, this gained impetus with the inauguration of "Programme on Nanomaterials: Science and Devices" in 2000 by the Department of Science and Technology (DST), Gol.

DST introduced special initiative to trigger and brace certain discontinuance projects leading paramount to concrete operations, products and technologies after acknowledging the predominance of nanomaterials and their immense overarching impact on technology (DST 2001).

In 2001-2002, the DST laid out an Expert Group on “Nanomaterials: Science and Devices”. The Government discerned the necessity to actuate a Nanomaterials Science and Technology Mission (NSTM) in the 10th Five Year Plan (2002-07) after taking into account the advancements in nanotechnology. A strategized framework was stipulated for supportive collaboration for futuristic approach in case of both research based and implementation aligned programmes in nanomaterials (DST 2001).

Documentation of the Tenth Five Year Plan (2002-2007) conformed the varied anticipation for operational programmes such as technology for bamboo products, drugs and pharmaceutical study, instrumental development including development of machinery and equipment, seismology, and also nano science and technology (GOI 2002).

Thereafter, the National Nanoscience and Nanotechnology Initiative (NSTI) was established in October, 2001 under the patronage of the Department of Science and Technology of the Ministry of Science. The stimulus for introducing NSTI in 2001 was to instil explorative infrastructure and publicise basic survey in nanoscience and nanotechnology. It mainly was observant on issues correlating with infrastructural betterment, standard research and application based programmes in nanomaterial including drugs, gene targeting and DNA chips. Nanotechnology was adjudged to be a trailblazing technology with influence in almost every facets of life. Enchanted by the never-ending possibilities of nanotechnological applications and in order to further enrich the visualisation of India 13 in nano science and technology, a Nano Science and Technology Mission (NSTM) was envisaged to provide desired momentum to developmental operations in this field (DST 2006).

The Eleventh Five-Year Plan (2007-2012) unremittingly designed project works to curate pose greater impact on socio-economic domain including nano material and nano devices in healthcare and disease. The generous Eleventh Five Year Plan Budget allocated an amount of Rs. 1000 crore for hypothecating the Nano Mission in 2007, with the objective to foster significant developmental changes and enhanced prospect in this arena.

Nano Science Advisory Group (NSAG) and the Nano Applications and Technology Advisory Group (NATAG) were engendered to inculcate structural development, public private partnerships, technological advancement centres and outlets, Human Resource Development, international collaborations, and Academia-Industry partnerships. In the Twelfth Five Year Plan (2012-2017), the government of India consented the prolongation of the Mission on Nano Science and Technology (Nano Mission) for its Phase-II.

The Council of Scientific and Industrial Research, also known as CSIR established 38 laboratories in India solely for extensive research work in Nanotechnology. Nanotechnology has certainly attained an indispensable stance in the Indian Economy and Scientific

Research Department and it is anticipated to gain the pinnacle of progress hence moulding India to emerge as one of role models on global map.[219]

8. Future Prospects

Nanotechnology, being the multi faceted line of discipline having its roots out stretched in almost every field in the proximity of mankind, holds a scope for improvement and development on regular terms. The efficient and dynamic nature of nanomaterials seek an improvement in application and technical aspect in its intricacies. The future research should be based on ways and techniques to diminish the risk factors related to the implications of nanomaterials such as - checking of permissible levels of nanoparticle dosage must be used within safety limit so that it has minimum or no adverse effects which can be achieved by concentration dependent studies in the natural soil system which can accurately reveal the non-toxic dose of various nanoparticles. A particular nanoparticle can have different effects on different kinds of plant. So elaborate study of the nanoparticle is required, which requires further investigation. Various nanoparticle based studies on plants have been made to apprise the varied regards under nanotechnology, but further comprehensive and extensive studies are in demand, to boost up plant growth and development with the least possible adverse outcomes.

9. Conclusion

The inter disciplinary field of nanotechnology has significant prospective in the study of plant sciences. Nanotechnology being both bane and boon, has multi faceted effect on plant growth and development. Studies conducted showed that nanoparticles have a variety of effects on plant physiology, morphology, response, interaction and overall plant growth evolution. It provides attributes to increase the plant productivity and resistance to many internal as well as external properties. Nanotechnology has helped improvising techniques in the field of agronomic sciences and has made it possible to minimize environmental pollution by producing chemical fertilizers and pesticides by utilizing nanoparticles and nano-capsules which has the ability to control or delay delivery in the intrusion of nanoparticles. With the aid of nano sciences, various plant diseases can be detected at an early stage so that tons of food produced can be protected from possible epidemic conditions which has enticed nanotechnologists for ways to safeguard the plants for bacteria, fungi and viral agents. It paves a way for the development of various techniques that take minimum time for detection, can give quick results, are elementary and precise and does not involve complicated techniques for the operation. Research related to nanotechnology involves the development and use of nanomaterials of various compounds. Nanomaterials can be prepared by various biological and biomimetic origins, with the help of varied techniques.

Critical and detailed analysis of appropriate information has unveiled that nanotechnology can have potential perspectives in the detection and diagnosis of a variety of plant diseases. Nanomaterials, microcapsules and nanotubes can efficiently carry high amount of active pesticide ingredients, chemicals which induce host resistance. Enzyme based biosensors which are covered with nanoparticles of various metals like Ti, Au, Ag, etc., may help in quick and accurate diagnosis of plant infection and would also help in detecting the remnant of pesticides. Development of nanopesticides can provide numerous advantages like: it can improve the solubility of pesticides, enhance the shelf-life, reduce the toxicity of pesticides, overcome the resistance of pesticides by the plants, etc,. In depth study and scrutiny of nanoparticle effect and working mechanism will make understanding of the nanomaterials more sound. Nanomaterials have an effect on the future prospect of mankind.

Conflict of Interest Declaration: On behalf of all authors, the corresponding author states that there is no conflict of interest.

References:

1. J. Safari and Z. Zarnegar (2014) Advanced drug delivery systems: nanotechnology of health design A review. *Journal of Saudi Chemical Society*: 85-99
2. O. P. Vilela Neto (2014) Intelligent computational nanotechnology: the role of computational intelligence in the development of nanoscience and nanotechnology. *Journal of Computational and Theoretical Nanoscience* 11(4) :928-944
3. V. Dutschk, T. Karapantsios, L. Liggieri, N. McMillan, R. Miller, V. M. Starov (2014) Smart and green interface from single bubbles/drops to industrial environmental and biomedical applications. *Advances in Colloid and Interface Science* 209: 109-126.
4. M. Al-Halafi (2014) Nanocarriers of nanotechnology in retinal disease. *Saudi Journal of Ophthalmology* 28(4): 304–309
5. P. Podsiadlo, A. K. Kaushik, E. M. Arruda et al (2007) Ultrastrong and stiff layered polymer nanocomposites. *Science*: 318, 80
6. Y. Vasquez, A. E. Henkes, J. Chris Bauer, R. E. Schaak (2008) Nanocrystal conversion chemistry: a unified and materials general strategy for the template-based synthesis of nanocrystalline solids. *Journal of Solid State Chemistry* 181(7):1509-1523
7. R. J. Mehta, Y. Zhang, C. Karthik (2012) A new class of doped nanobulk high-figure-of-merit thermoelectrics by scalable bottom-up assembly. *Nature Materials* 11(3):233-240
8. X. Lang, A. Hirata, T. Fujita, M. Chen (2011) Nanoporous metal/oxide hybrid electrodes for electrochemical supercapacitors. *Nature Nanotechnology* 6(4):232-6
9. L. Rizzello and P. P. Pompa (2014) Nanosilver-based antibacterial drugs and devices: mechanisms, methodological drawbacks, and guidelines. *Chemical Society Reviews* 6(4):232-6

10. D. Astruc (2012) Electron-transfer processes in dendrimers and their implication in biology, catalysis, sensing and nanotechnology. *Nature Chemistry*: 4(4):255-67
11. Syed Salman Abbas, Mohd Haneef, Mohtashim Lohani, Heena Tabassum, Ahamad Faiz Khan (2016) Nanomaterials Used As A Plants Growth Enhancer: An Update. *International Journal of Pharma Research & Review* 5:17-23.
12. Baughman RH, Zakhidov AA, and W. A. de Heer (2002) Carbon nanotubes—the route toward application. *Science* 297(5582):787–792.
13. Lang X, Hirata A, Fujita T, Chen M (2011) Nanoporous Metal/Oxide Hybrid Electrodes For Electrochemical Supercapacitors. *Nature Nanotechnology* 6(4):232– 236.
14. Rizzello L, Pompa PP (2014) Nanosilver-based antibacterial drugs and devices: mechanisms, methodological drawbacks, and guidelines. *Chemical Society Reviews* 43(5):1501–1518.
15. Yang K, Ma Y (2010) Computer simulation of the translocation of nanoparticles with different shapes across a lipid bilayer. *Nature Nanotechnology* 5(8):579– 583
16. Lang X, Hirata A, Fujita T, Chen M (2011) Nanoporous Metal/Oxide Hybrid Electrodes For Electrochemical Supercapacitors. *Nature Nanotechnology* 6(4):232– 239
17. Rizzello L, Pompa PP (2014) Nanosilver-based antibacterial drugs and devices: mechanisms, methodological drawbacks, and guidelines. *Chemical Society Reviews* 43(5):1501–1518.
18. Ljubimova JY, Holler E (2012) Biocompatible nanopolymers: the next generation of breast cancer treatment? *Nanomedicine* 7(10):1467–1470
19. D. Astruc (2012) Electron-transfer processes in dendrimers and their implication in biology, catalysis, sensing and nanotechnology. *Nature Chemistry*: 4(4):255-67
20. Zhang L, Fang M (2010) Nanomaterials in pollution trace detection and environmental improvement. *Nano Today* 5(2):128–142.
21. Oberdörster G, Oberdörster E, Oberdörster J (2005) Nanotoxicology: An emerging discipline evolving from studies of ultrafineparticles. *Environ Health Perspect* 113:823–839.
22. Maynard AD, Aitken RJ, Butz T, Colvin V, Donaldson K, Oberdörster G, Philbert MA, Ryan J, Seaton A, Stone V et al (2006). Safe handling of nanotechnology *Nature* 444:267–269
23. Orive G, Anitua E, Pedraz J, Emerich D (2009) Biomaterials for promoting brain protection, repair and regeneration. *Nat. Rev. Neurosci* 10(9):682-692.
24. Adair J, Parette M, Erhan I, Kester M.(2010) Nanoparticulate alternatives for drug delivery. *ACS Nano* 4(9):4967-70
25. Ratner B, Hoffman A, Schoen F, Lemons J (2014) *Biomaterials Science: An Introduction to Materials in Medicine*. Third Edit. Canada: Academic Press
26. Akbarzadeh A, Rezaei-Sadabady R, Davaran S, et al (2013) Liposome: classification, preparation, and applications. *Nanoscale Res. Lett.* 8(102):1-9.
27. Reis C, Neufeld R, Ribeiro A, Veiga F. Nanoencapsulation I (2006) Methods for preparation of drug-loaded polymeric nanoparticles. *Nanomedicine* 2(1):8-21.
28. Zhang L, Chan J, Gu F (2008) Self – assembled lipid polymer hybrid Nanoparticles : a robust drug delivery platform. *ACS Nano* 2(8):1696-702

29. Hu C-M, Zhang L, Aryal S, Cheung C, Fang R, Zhang L (2011) Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. *Proc. Natl. Acad. Sci. U. S. A.* 108(27):10980-10985.
30. Hadinoto K, Sundaresan A, Cheow W (2013) Lipid-polymer hybrid nanoparticles as a new generation therapeutic delivery platform: a review. *Eur. J. Pharm. Biopharm* 14:1937-1952
31. Chen Z, Ma L, Liu Y, Chen C (2012) Applications of functionalized fullerenes in tumor theranostics. *Theranostics* 2(3):238-250.
32. Ma H, Liang X-J (2010) Fullerenes as unique nanopharmaceuticals for disease treatment. *Sci. China Chem* 53(11):2233-2240
33. Chakravarty S, Kivelson S.(1991) Superconductivity of foped fullerenes. *Europhys. Lett.* 16(8):751-756
34. Klimov VI (2007) Spectral and dynamical properties of multiexcitons in semiconductor nanocrystals. *Annu Rev Phys Chem* 58:635–673.
35. Valizadeh A, Mikaeili H, Samiei M, Farkhani SM, Zarghami N, Kouhi M, Akbarzadeh A, Davaran S (2012) Quantum dots: synthesis, bioapplications, and toxicity. *Nanoscale Res Lett* 7(1):480.
36. Bruchez M Jr, Moronne M, Gin P, Weiss S, Alivisatos AP (1998) Semiconductor nanocrystals as fluorescent biological labels. *Science* 281(5385):2013-2016
37. Nair R, Poulouse AC, Nagaoka Y, Yoshida Y, Maekawa T, Kumar DS (2011) Uptake of FITC labeled silica nanoparticles and quantum dots by rice seedlings: effects on seed germination and their potential as biolabels for plants. *J Fluoresc* 21(6):2057–2068.
38. Hoshino A, Fujioka K, Oku T, Suga M, Sasaki YF, Ohta T, Yasuhara M, Suzuki K, Yamamoto K (2004) Physicochemical properties and cellular toxicity of nanocrystal quantum dots depend on their surface modification. *Nano Lett* 4(11):2163–2169.
39. Shiohara A, Hoshino A, Hanaki K, Suzuki K, Yamamoto K (2004) On the cytotoxicity caused by quantum dots. *Microbiol Immunol* 48(9):669–675
40. Alimohammadi M, Xu Y, Wang DY, Biris AS, Khodakovskaya MV (2011) Physiological responses induced in tomato plants by a two-component nanostructural system composed of carbon nanotubes conjugated with quantum dots and its in vivo multimodal detection. *Nanotechnology* 22(29):295101
41. Navarro DA, Bisson MA, Aga DS (2012) Investigating uptake of water-dispersible CdSe/ZnS quantum dot nanoparticles by *Arabidopsis thaliana* plants. *J Hazard Mater* 211–212:427– 435.
42. Stefaniak, Aleksandr B. (2017). *Principal Metrics and Instrumentation for Characterization of Engineered Nanomaterials.*
43. P. Heera, S. Shanmugam (2015) Nanoparticle Characterization and Application: An Overview. *Int.J.Curr.Microbiol.App.Sci* 4(8):379-386
44. Joshi ,Bhattacharya, Ali (2008) Characterisation techniques for nanotechnology in textiles. *Indian journal of fibre and textile research* 33:304-317

45. C. M. Rico, S. Majumdar, M. Duarte-Gardea, J. R. Peralta-Videa, J. L. Gardea-Torresdey (2011) Interaction of Nanoparticles with edible plants and their possible implications in the food chain. *Journal of Agricultural and Food Chemistry* 59(8): 3485–3498
46. Lin S, Reppert J, Hu Q, Hudson JS, Reid ML, Ratnikova TA, Rao AM, Luo H, Ke PC (2009) Uptake, translocation, and transmission of carbon nanomaterials in rice plants *Small* 5: 1128–1132.
47. Hall JL & Williams LE (2003) Transition metal transporters in plants. *J Exp Bot* 54:2601–2613.
48. Whitby M & Quirke N (2007) Fluid flow in carbon nanotubes and nanopipes. *Nat Nanotechnol* 2.
49. Lin D, Xing B (2008) Root uptake and phytotoxicity of ZnO nanoparticles. *Environ Sci Technol* 42:5580–5585.
50. Birbaum K, Brogiolli R, Schellenberg M, Martinoia E, Stark WJ, Gunther D, Limbach L No evidence for cerium dioxide nanoparticle translocation in maize plants. *Environ Sci Technol* 44(22): 8718-8723
51. Zhu H, Han J, Xiao JQ, Jin Y (2008) Uptake, translocation, and accumulation of manufactured iron oxide by pumpkin plants. *J Environ Monit* 10: 713–717.
52. Lee WM, An YJ, Yoon H, Kweon HS (2008) Toxicity and bioavailability of copper nanoparticles to the terrestrial plants mung bean (*Phaseolus radiatus*) and wheat (*Triticum aestivum*): plant agar test for water-insoluble nanoparticles. *Nanomat Environ* 27: 1915–1921.
53. Stampoulis D, Sinha SK, White JC (2009) Assay-dependent phytotoxicity of nanoparticles to plants. *Environ Sci Technol* 43: 9473–9479.
54. Navarro E, Baun A, Behra R, Hartmann NB, Filser J, Miao AJ, Quigg A, Santschi PH, Sigg L (2008) Environmental behaviour and ecotoxicity of engineered nanoparticles to algae, plants, and fungi. *Ecotoxicology* 17: 372–386.
55. Judy JD, Unrine JM, Bertsch PM (2011) Evidence for biomagnification of gold nanoparticles within a terrestrial food chain. *Environ Sci Technol* 45: 776–781.
56. Grieger KD, Hansen SF, Baun A (2009) The known unknowns of nanomaterials: describing and characterizing uncertainty within environmental, health and safety risks. *Nanotoxicology* 3(3): 222–233.
57. Zhang L, Fang M, (2010) Nanomaterials in pollution trace detection and environmental improvement. *Nano Today* 5(2):128–142.
58. Alexis DO, Tyronne M, Joseph C, Indy H, Barbara HH (2009) Nanotoxicology: characterizing the scientific literature. *Journal of Nanoparticle Research* 11(2): 251–257
59. Baughman RH, Zakhidov AA, de Heer WA (2002) Carbon nanotubes—the route toward applications. *Science* 297(5582): 787–792.
60. R. De La Torre-Roche, Hawthorne J, Deng Y et al.(2013) Multiwalled carbon nanotubes and C60 fullerenes differentially impact the accumulation of weathered pesticides in four agricultural plants. *Environmental Science and Technology* 47(21): 12539–12547.
61. Liu Q, Zhang X, Zhao Y (2013) Fullerene-induced increase of glycosyl residue on living plant cell wall. *Environmental Science and Technology* 47(13): 7490–7498.
62. Santos SMA, Dinis AM, Rodrigues DMF, Peixoto F, RA. Videira, A. S. Jurado (2013), Studies on the toxicity of an aqueous suspension of C60 nanoparticles

- using a bacterium (gen. *Bacillus*) and an aquatic plant (*Lemna gibba*) as in vitro model systems. *Aquatic Toxicology* 142–143:347–354.
63. De La Torre-Roche R, Hawthorne J, Deng Y (2012) Fullerene enhanced accumulation of p, p-DDE in agricultural crop species. *Environmental Science and Technology* 46(17):9315–9323.
 64. Gao J, Wang Y, Folta KM (2011) Polyhydroxy fullerenes (fullerols or fullerlenols): beneficial effects on growth and lifespan in diverse biological models. *PLoS ONE* 6(5): e19976
 65. Liu Q, Zhao Y, Wan Y (2010) Study of the inhibitory effect of water-soluble fullerenes on plant growth at the cellular level. *CS Nano* 4(10): 5743–5748.
 66. Lin Y, Rao AM, Sadanadan B, Kenik EA, Sun YP (2002) Functionalizing multiple-walled carbon nanotubes with aminopolymers. *Journal of Physical Chemistry B* 106(6): 1294–1298.
 67. Muller J, Huaux , Moreau N et al. (2005) Respiratory toxicity of multi-wall carbon nanotubes. *Toxicology and Applied Pharmacology* 207(3):221–231.
 68. Kong H, Gao C, Yan D (2004) Controlled functionalization of multiwalled carbon nanotubes by in situ atom transfer radical polymerization. *Journal of the American Chemical Society* 126(2):412–413
 69. Li HJ, Lu WG, Li JJ, Bai XD, Gu CZ (2005) Multichannel ballistic transport in multiwall carbon nanotubes. *Physical Review Letters* 95(8): 086601
 70. Chekin F, Bagheri S, Arof AK, Hamid SBA (2012) Preparation and characterization of Ni(II)/ and carbon nanotube composite modified electrode and application for carbohydrates electrocatalytic oxidation. *Journal of Solid State Electrochemistry* 16(10): 3245–3251.
 71. Aslani, F., Bagheri, S., Muhd Julkapli, N., Juraimi, A. S., Hashemi, F. S., & Baghdadi, A. (2014). Effects of engineered nanomaterials on plants growth: an overview. *The Scientific World Journal*, 2014:641759.
 72. Smirnova E., Gusev A, Zaytseva O et al (2012) Uptake and accumulation of multiwalled carbon nanotubes change themorphometric and biochemical characteristics of *Onobrychis arenaria* seedlings. *Frontiers of Chemical Science and Engineering* 6(2):132–138
 73. X.-M. Tan, Fugetsu B (2007) Multi-walled carbon nanotubes interact with cultured rice cells: evidence of a self-defense response. *Journal of Biomedical Nanotechnology* 3(3): 285–288.
 74. Lou JC, Jung MJ, Yang HW, Han JY, Huang WH (2011) Removal of dissolved organic matter (DOM) from raw water by single-walled carbon nanotubes (SWCNTs). *Journal of Environmental Science and Health—Part A Toxic/Hazardous Substances and Environmental Engineering* 46(12):1357–1365.
 75. Yuan H, Hu S, Huang P et al.(2011) Single walled carbon nanotubes exhibit dual-phase regulation to exposed *Arabidopsis mesophyll* cells. *Nanoscale Research Letters* 6(1):1–9.
 76. Soylak M, Unsal YE (2010) Chromium and iron determinations in food and herbal plant samples by atomic absorption spectrometry after solid phase extraction on single-walled carbon nanotubes (SWCNTs) disk. *Food and Chemical Toxicology* 48(6):1511–1515.
 77. M. S. Eva, B. Anders, K. Matthias, T. Stefan (2009) Insignificant acute toxicity of TiO₂ nanoparticles to willow trees. *Journal of Soils and Sediments* 9(1): 46–53

78. Feizi H, Rezvani Moghaddam P, Shahtahmassebi N, A. Fotovat (2012) Impact of bulk and nanosized titaniumdioxide (TiO₂) on wheat seed germination and seedling growth. *Biological Trace Element Research* 146(1):101–106.
79. Mingfang Q, Yufeng L, Tianlai L (2013) Nano-TiO₂ improve the photosynthesis of tomato leaves under mild heat stress, biological trace element research. *Biological Trace Element Research* 156(1): 323–328.
80. Uhram S, Minjoo S, Gisuk L, Jinkyu R, Younghun K, Eun JL (2013) Functional analysis of TiO₂ nanoparticle toxicity in three plant species. *Biological Trace Element Research* 155(1): 93–103.
81. Zheng L, Hong, Lu S, Liu C (2005) Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biological Trace Element Research* 104(1): 83–91.
82. Yang F, Hong F, You W et al.(2006) Influences of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. *Biological Trace Element Research* 110(2): 179–190.
83. Castiglione M, Giorgetti L, Geri C, Cremonini R (2011) The effects of nano-TiO₂ on seed germination, development and mitosis of root tip cells of *Vicia narbonensis* L. and *Zeamays* L. *Journal of Nanoparticle Research* 13(6): 2443–2449.
84. Kurepa J, Paunesku T, S. Vogt et al (2010) Uptake and distribution of ultrasmall anatase TiO₂ alizarin red S nanoconjugates in *arabidopsis thaliana*. *Nano Letters* 10(7): 2296–2302.
85. Wang S, Kurepa J, Smalle JA (2011) Ultra-small TiO₂ nanoparticles disrupt microtubular networks in *Arabidopsis thaliana*. *Plant, Cell & Environment* 34(5):811–820.
86. Eduardo Corredor, Pilar S Testillano, María-José Coronado, Pablo González-Melendi, Rodrigo Fernández-Pacheco, Clara Marquina, M Ricardo Ibarra, Jesús M de la Fuente, Diego Rubiales, Alejandro Pérezde- Luque, María-Carmen Risueño et al (2009) Nanoparticle penetration and transport in living pumpkin plants: in situ subcellular identification. *BMC Plant Biology* 9:45
87. Ju-Nam Y, Lead JR (2008) Manufactured nanoparticles: an overview of their chemistry, interactions and potential environmental implications. *Sci Total Environ.* 400: 396-414
88. Aitken RJ, Chaudhry MQ, Boxall ABA, Hull M.(2006) Manufacture and use of nanomaterials: current status in the UK and global trends. *Occup Med.* 56(5):300-6.
89. Garcia M, Forbe T, Gonzalez E (2010) Potential applications of nanotechnology in the agro-food sector. *Ciencia Tecnol Aliment* vol.30, n.3, pp.573-581
90. Milne W.I, Teo K.B.K, Amaratunga G.A.J, Legagneux P, Gangloff L, Schnell J.P, Semet V, Binh, V.T., Groening, O. Carbon nanotubes as field emission sources. *J. Mater. Chem.* 14:933-943
91. Nair R, Varghese S.H, Nair B.G, Maekawa T, Yoshida Y, Kumar D.S. (2010) Nanoparticulate material delivery to plants. *Plant Sci.* 179: 154–163

92. J.M. Green, G.B. Beestman (2007) Recently patented and commercialized formulation and adjuvant technology. *Crop Prot.* 26(3):320-327
93. A.L. Boehm, I. Martinon, R. Zerrouk, E. Rump, H. Fessi (2003) Nanoprecipitation technique for the encapsulation of agrochemical active ingredients. *J Microencapsul.* 20(4):433-41
94. Jie Yang, Weidong Cao & Yukui Rui (2017) Interactions between nanoparticles and plants: phytotoxicity and defense mechanisms. *Journal of Plant Interactions* 59(8): 3485-3498
95. Parvin Begum, Refi Ikhtiari, Bunshi Fugetsu et al (2014) Potential Impact of Multi-Walled Carbon Nanotubes Exposure to the Seedling Stage of Selected Plant Species. *Nanomaterials* 4: 203-221
96. Archana Yadav, T.Theivasanthi, P.K.Paul and K.C. Upadhyay (2015) Extracellular biosynthesis of silver nanoparticles from Plant Growth Promoting Rhizobacteria *Pseudomonas* sp. *Int.J.Curr.Microbiol.App.Sci* 4(8): 1057-1068
97. Bangale M.S, Mitkare S.S, Gattani S.G, Sakarkar D.M. (2012) Recent nanotechnological aspects in cosmetics and dermatological preparations. *Int. J. Pharm. Pharm. Sci.* 4(2):88-97
98. Vance M.E, Kuiken T, Vejerano E.P, McGinnis S.P, Hochella M.F, Hull D.R. (2015) Nanotechnology in the real world: Redeveloping the nanomaterial consumer products inventory. *Beilstein J. Nanotechnol.* 6:1769-80
99. Jacoby R, Peukert M, Succurro A, Koprivova A, Kopriva S. The Role of Soil Microorganisms in Plant Mineral Nutrition—Current Knowledge and Future Directions. *Front. Plant Sci.* 8: 1617
100. Cavassin E.D, de Figueiredo L.F.P, Otoch J.P, Seckler M.M, de Oliveira R.A, Franco F.F, Marangoni V.S, Zucolotto V, Levin A.S.S, Costa S.F (2015) Comparison of methods to detect the in vitro activity of silver nanoparticles (AgNP) against multidrug resistant bacteria. *J. Nanobiotechnol.* 8(5): 279.
101. Radzig M.A., Nadochenko V.A, Koksharova O.A, Kiwi J, Lipasova V.A, Khmel I.A (2013) Antibacterial effects of silver nanoparticles on gram-negative bacteria: Influence on the growth and biofilms formation, mechanisms of action. *Colloids Surf. B Biointerfaces* 102:300-306
102. Sondi I, Salopek-Sondi B. (2004) Silver nanoparticles as antimicrobial agent: A case study on *E. coli* as a model for Gram-negative bacteria. *J. Colloid Interface Sci.* 275(1):177-182
103. Stanković A, Dimitrijević S, Uskoković D (2013) Influence of size scale and morphology on antibacterial properties of ZnO powders hydrothermally synthesized using different surface stabilizing agents. *Colloids Surf. B Biointerfaces* 102:21-28
104. Frenk S, Ben-Moshe T, Dror I, Berkowitz B, Minz D (2013) Effect of Metal Oxide Nanoparticles on Microbial Community Structure and Function in Two Different Soil Types. *PLoS ONE* 8(12): e84441
105. Colman B.P, Arnaout C.L, Anciaux S, Gunsch C.K, Hochella M.F, Kim B, Lowry G.V, McGill B.M, Reinsch B.C, Richardson C.J (2013) Low Concentrations

- of Silver Nanoparticles in Biosolids Cause Adverse Ecosystem Responses under Realistic Field Scenario. PLoS ONE 27(3): 482-490
106. Ge Y, Schimel, J.P, Holden P.A. (2011) Evidence for negative effects of TiO₂ and ZnO nanoparticles on soil bacterial communities. *Env. Sci. Technol.* 45(4):1659-64
 107. Sangeeta Chavan, Vigneshwaran Nandanathangam (2019) Effects of Nanoparticles on Plant Growth-Promoting Bacteria in Indian Agricultural Soil. *Agronomy* 9(3):140
 108. Giuliano Degrassi, Iris Bertani, Giulia Devescovi, Alberto Fabrizi, Antonietta Gatti, Vittorio Venturi, (2012) Response Of Plant-Bacteria Interaction Models To Nanoparticles. *Qualità ambientale.* 8: 39-50
 - Nel A, Xia T, Mädler L, Li N (2006) Toxic potential of materials at the nanolevel. *Science* 311(5761):622-627
 109. Nel, A., Xia, T., Mädler, L. Li, N (2006) Toxic potential of materials at the nanolevel. *Science* 311(5761):622-627
 110. Auffan M, Achouak W, Rose J, Roncato M.A, Chanéac C, Waite D.T, Masion A, Woicik J.C, Wiesner M.R, Bottero J.Y.(2008) Relation between the redox state of iron-based nanoparticles and their cytotoxicity toward Escherichia coli. *Environ. Sci. Technol.* 42(17):6730-6735
 111. Foley S, Crowley C, Smaih M, Bonfils C, Erlanger B.F, Seta P, Larroque C. (2002) Cellular localisation of a water-soluble fullerene derivative. *Biochem. Biophys. Res. Commun.* 294(1):116-119
 112. Kamat J.P, Devasagayam T.P, Priyadarsini K.I, Mohan H. (2000) Reactive oxygen species mediated membrane damage induced by fullerene derivatives and its possible biological implications. *Toxicology* 155(1-3):55-61
 113. Gopinath P, Gogoi S.K, Sanpui P, Paul A, Chattopadhyay A, Ghosh S.S (2010) Signaling gene cascade in silver nanoparticle induced apoptosis. *Colloids Surf. B* 77(2):240-245
 114. Slomberg D.L, Schoenfisch M.H. (2012) Silica nanoparticle phytotoxicity to Arabidopsis thaliana. *Environ. Sci. Technol.* 46(18):10247-10254
 115. Begum P, Fugetsu B. (2012) Phytotoxicity of multi-walled carbon nanotubes on red spinach (*Amaranthus tricolor* L) and the role of ascorbic acid as an antioxidant. *J. Hazard. Mater.* 243:212-222
 116. Kumar V, Yadav SK (2009) Plant-mediated synthesis of silver and gold nanoparticles and their applications. *J Chem Technol Biotechnol* 84(2): 151-157
 117. Sharma VK, Yngard RA, Lin (2009) Silver nanoparticles: Green synthesis and their antimicrobial activities. *Adv Colloid Interface Sci.* 145(1-2):83-96
 118. Siddiqui MH, Al-Whaibi MH, Faisal M, Al Sahli AA (2014) Nanosilicon dioxide mitigates the adverse effects of salt stress on Cucurbita pepo L. *Environ Toxicol Chem* 33(11):2429-2437
 119. Nair PMG, Chung IM. (2014) Impact of copper oxide nanoparticles exposure on Arabidopsis thaliana growth, root system development, root lignification, and molecular level changes. *Environ Sci Pollut Res* 21(22):12709-12722

120. Prakash MG, Chung IM. (2016) Determination of zinc oxide nanoparticles toxicity in root growth in wheat (*Triticum aestivum* L.) seedlings. *Acta Biol Hung* 67(3):286-296.
121. Adams J, Wright M, Wagner H (2016) Cu from dissolution of CuO nanoparticles signals changes in root morphology. *Plant Physiol Biochem* 110:108-117
122. Dimkpa CO, Calder A, Britt DW (2011) Responses of a soil bacterium, *Pseudomonas chlororaphis* O6 to commercial metal oxide nanoparticles compared with responses to metal ions. *Environ Pollut* 159(7):1749-1756
123. Patra, H. P Adhikari, Shyamal Kumar, Kunar Subrata (2016) Groundwater Prospecting and Management. *Hydrogeology*
124. Josef JAMPÍLEK, Katarína KRÁĽOVÁ, *ECOL CHEM ENG S* (2015) Application Of Nanotechnology In Agriculture and Food Industry. Its Prospects And Risks 22(3):321-361
125. Melek Ekinci, Atilla Dursun, Ertan Yıldırım, Fazilet Parlakova (2014) Effects of Nanotechnology liquid fertilizers on plant growth and yield of cucumber (*Cucumis sativus* L.) *Acta Sci. Pol.* 13(3):135-141.
126. Bhanushali Mansi P1, Jaybhaye Sandesh V2, Gutte Arun V3 (2017) Copper nanoparticles using Onion (*Allium cepa*) extract and their application in plant growth. *Int. J. of Life Sciences* 5 (4): 661-666
127. Y. Qi, K. Lian, Q. Wu, Y. Li, M. Danzy, R. Menard, K. L. Chin, D. Collins, F. Oliveria, K. Klepzig (2011) Nanotechnology application in forest protection. In *Proceedings of the TAPPI International Conference on Nanotechnology for Renewable Materials*, Washington, DC, USA : 271–301.
128. Lee, W., An, Y., Yoon, H., Kweon, H.(2008) Toxicity and bioavailability of copper nanoparticles to the terrestrial plants mung bean (*Phaseolus radiatus*) and wheat (*Triticum aestivum*): Plant uptake for water insoluble nanoparticles. *Environ. Toxicol. Chem.* 27: 1915–1921.
129. Husen, A.; Siddiqi, K.S. (2014) Phytosynthesis of nanoparticles: Concept, controversy and application. *Nanoscale Res. Lett.* Na 9(9):229
130. Juhel G, Batisse E, Hugues Q, Daly D, Van pelt F. N, O Halloran J. ,Jansen M. A.(2011) Alumina nanoparticles enhance growth of *Lemna minor*. *Aquat Toxicol.* 105(3–4): 328–326
131. Ernest V., Shiney P. J., Mukherjee A., Chandrasekaran N. (2012) Silver nanoparticles: a potential nanocatalyst for the rapid degradation of starch hydrolysis by amylase. *Carbohydr. Res.*, 352(60–64):22.
132. Chen Chen, Xiaodong Sun, Wu Pan, Yi Hou, Rui Liu, Xin Jiang, Li Zhang (2018) Graphene Oxide-Templated Synthesis of Hydroxyapatite Nanowhiskers To Improve the Mechanical and Osteoblastic Performance of Poly(lactic acid) for Bone Tissue Regeneration. *ACS Sustainable Chem. Eng.* 6(3):3862-3869
133. Raja Muthuramalingam Thangavelu (2019) Effect of deoxycholate capped silver nanoparticles in seed dormancy breaking of *Withania somnifera*
134. R. C. Monica and R. Cremonini (2009) Nanoparticles and higher plants," *Caryologia*, (62) : 161-165

135. M. Sharon, A. K. Choudhary, and R. Kumar (2010) Nanotechnology in agricultural diseases and food safety. *Journal of Phytology*. 2(4): 83-92
136. Seyed Saeid Hojjat, Hamidreza Hojjat (2015) Effect of Nano Silver on Seed Germination and Seedling Growth in Fenugreek Seed. *International Journal of Food Engineering* 1(2):106-110
137. Mahmoodzadeh H, Nabavi M, Kashefi H (2013) Effect of nanoscale titanium dioxide particles on the germination and growth of canola (*Brassica napus*). *J Ornamental Hortic Plants* 3:25–32
138. Jaberzadeh A, Moaveni P, Moghadam HRT, Zahedi H (2013) Influence of bulk and nanoparticles titanium foliar application on some agronomic traits, seed gluten and starch contents of wheat subjected to water deficit stress. *Not Bot Horti Agrobi* 41:201–207
139. V. Mishra, R.K. Mishra, A. Dikshit, A.C. Panday (2014) Interactions of nanoparticles with plants: an emerging prospective in the agriculture industry. In: Ahmad P, Rasool S (eds) *Emerging technologies and management of crop stress tolerance: biological techniques (1)* : 159–180
140. Yang F, Hong F, You W, Liu C, Gao F, Wu C, Yang P (2006) Influence of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. *Biol Trace Elem Res* 110(2):179–190
141. de la Rosa G, Lopez-Moreno ML, de Haro D, Botez CE, Peralta-Videa JR, Gardea-Torresdey JL (2013) Effects of ZnO nanoparticles in alfalfa, tomato, and cucumber at the germination stage: root development and X-ray absorption spectroscopy studies. *Pure Appl Chem* 85(12):2161–2174
142. F. Hong, J. Zhou, C. Liu, F. Yang, C. Wu, L. Zheng, P. Yang (2005) Effects of nano-TiO₂ on photochemical reaction of chloroplasts of Spinach. *Biological Trace Element Research* 105:269-279
143. X. M. Liu, F. D. Zhang, et al.(2005) Effects of nano-ferric oxide on the growth and nutrients absorption of peanut. *Plant Nutrition and Fertilizer Sci.*11:14-18
144. Shah V, Belozeroval I (2009) Influence of metal nanoparticles on the soil microbial community and germination of lettuce seeds. *Water Air Soil Pollut* 197:143–148
145. Barrena R, Casals E, Colón J, Font X, Sánchez A, Puntès V (2009) Evaluation of the ecotoxicity of model nanoparticles. *Chemosphere* 75(7):850–857
146. Arora S, Sharma P, Kumar S, Nayan R, Khanna PK, Zaidi MGH (2012) Gold-nanoparticle induced enhancement in growth and seed yield of *Brassica juncea*. *Plant Growth Regul* 66:303–310
147. Savithamma N, Ankanna S, Bhumi G (2012) Effect of nanoparticles on seed germination and seedling growth of *Boswellia ovalifoliolata* an endemic and endangered medicinal tree taxon. *Nano Vision* 2:61–68
148. Gopinath K, Gowri S, Karthika V, Arumugam A (2014) Green synthesis of gold nanoparticles from fruit extract of *Terminalia arjuna*, for the enhanced seed germination activity of *Gloriosa superba*. *J Nanostruct Chem* 4: 1–11

149. Christou P, McCabe DE, Swain WF (1988) Stable transformation of soybean callus by DNAcoated gold particles. *Plant Physiol* 87:671–674
150. Kumar V, Guleria P, Kumar V, Yadav SK (2013) Gold nanoparticle exposure induces growth and yield enhancement in *Arabidopsis thaliana*. *Sci Total Environ* 461:462–468
151. Richmond KE, Sussman M (2003) Got silicon? The non-essential beneficial plant nutrient. *Current Opinion in Plant Biology* 6: 268 272..
152. Ma JF, Yamaji N (2006) Silicon uptake and accumulation in higher plants. *Trends in Plant Science* 11: 392 397.
153. Zhu J (2003) Regulation homeostasis under salt stress. *Current Opinion in cell Biology* 6(5): 141 145.
154. Shi X H, Zhang CC, Wang H , Zhang FS (2005) Effect of Si on the distribution of Cd in rice seedling. *Plant and Soil* 273: 53-60.
155. Gunes A, Inal A, Bagic EG (2007) Silicon mediated changes of some physiological and enzymatic parameters symptomatic for oxidative stress in spinach and tomato grown in sodic – B toxic soil. *Plant and Soil* 290: 103 114.
156. Shi G, Cai Q, Liu C & Wu L (2010) Silicon alleviates cadmium toxicity in peanut plants in relation to cadmium distribution and stimulation of antioxidative enzymes. *Plant Growth Regulation* 61: 45-52
157. Rajoriya A, Tripathy SS & Gupta N (2015) In vitro antioxidant activity of selected *Ganoderma* species found in Odisha, India. *Tropical Plant Research* 2(2): 72 –77
158. B. Kisan, H. Shruthi, H. Sharanagouda, S.B. Revanappa, N.K. Pramod (2015) Effect of Nano-Zinc Oxide on the Leaf Physical and Nutritional Quality of Spinach, *Agrotechnology*
159. Prasad TNVKV, Sudhakar P, Sreenivasulu Y, Latha P, Munaswamy V, Reddy KR et al.(2012) Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. *J Plant Nutr.* 35(6):905-927.
160. M. Sedghi, M. Hadi, S.G. Toluie (2013) Effect of nano zinc oxide on the germination of soybean seeds under drought stress. *Ann West Uni Timis, oarsa ser Biol XVI (2) : 73-78*
161. Ramesh M, Palanisamy K, Babu K, Sharma NK (2014) Effects of bulk & nanotitanium dioxide and zinc oxide on physio-morphological changes in *Triticum aestivum* Linn. *J Glob Biosci.* 3:415-422
162. Raskar SV, Laware SL.(2014) Effect of zinc oxide nanoparticle on cytology and seed germination in onion. *Int. J. Curr.Microbiol. App. Sci.* 3(2):467-473
163. Ambily Elizabeth, Vijay Bahadur, Pragathi Misra, Vipin Mashhi Prasad and T Thomas (2017) Effect of different concentrations of iron oxide and zinc oxide nanoparticles on growth and yield of carrot (*Daucus carota* L.) *Journal of Pharmacognosy and Phytochemistry* 6(4): 1266-1269
164. Guadalupe de la Rosa, Martha Laura López-Moreno, David de Haro, Cristian E. Botez, José R. Peralta-Videa, Jorge L. Gardea-Torresdey (2013) Effects of ZnO nanoparticles in alfalfa, tomato, and cucumber at the germination stage: Root

- development and X-ray absorption spectroscopy studies. *Pure Appl. Chem.* 85(12): 2161–2174
165. Khodakovskaya M, Dervishi E, Mahmood M, Xu Y, Li Z, Watanabe F, Biris AS,(2009) Carbon nanotubes are able to penetrate plant seed coat and dramatically affect seed germination and plant growth. *ACS Nano*, 3: 3221–3227.
166. Srinivasan C, Saraswathi R (2010) Nano-agriculture-carbon nanotubes enhance tomato seed germination and plant growth. *Curr Sci* 99: 273–275.
167. Villagarcia H, Dervishi E, Silva K, Biris AS, Khodakovskaya MV (2012) Surface chemistry of carbon nanotubes impacts the growth and expression of water channel protein in tomato plants. *Small*, 8:2328–2334.
168. Tiwari DK, Dasgupta-Schubert N, Villaseñor-Cendejas LM, Villegas J, Carreto-Montoya L, Borjas-García SE (2014) Interfacing carbon nanotubes (CNT) with plants: Enhancement of growth, water and ionic nutrient uptake in maize (*Zea Mays*) and implications for nanoagriculture. *Appl Nanosci* 4:577–591.
169. Lahiani MH, Dervishi E, Chen J, Nima Z, Gaume A, Biris AS, Khodakovskaya MV (2013) Impact of carbon nanotube exposure to seeds of valuable crops. *ACS Appl Mater*, 5:7965–7973.
170. Wang X, Han H, Liu X, Gu X, Chen K , Lu D,(2012) Multi-walled carbon nanotubes can enhance root elongation of wheat (*Triticum aestivum*) plants. *J Nanopart Res*, 14(6): 1–10.
171. S. Tripathi, S. Sarkar (2014) Influence of water soluble carbon dots on the growth of wheat plant. *Appl Nanosci.* (5) : 609–616
172. Lin D , Xing B,(2007) Phytotoxicity of nanoparticles: inhibition of seed germination and root growth. *Environ. Pollut* 150:243–250.
173. Canas JE, Long M, Nations S, Vadan R, Dai L, Luo M et al,(2008) Effects of functionalized and nonfunctionalized single-walled carbon nanotubes on root elongation of select crop species. *Environ. Toxicol. Chem* 27:1922–1931.
174. L.R. Pokhrel, B. Dubey (2013) Evaluation of developmental responses of two crop plants exposed to silver and zinc oxide nanoparticles, *Sci.Tot.Environ.* 321–332
175. Arora S, Sharma P, Kumar S, Nayan R, Khanna PK, Zaidi MGH (2012) Gold-nanoparticle induced enhancement in growth and seed yield of *Brassica juncea*. *Plant Growth Regul* 66: 303–310.
176. Gopinath K, Gowri S, Karthika V, Arumugam A,(2014) Green synthesis of gold nanoparticles from fruit extract of *Terminalia arjuna*, for the enhanced seed germination activity of *Gloriosa superba*. *J Nanostruct Chem* 4:1–11.
177. V. Kumar, P. Guleria , V. Kumar, S.K. Yadav (2013) Gold nanoparticle exposure induces growth and yield enhancement in *Arabidopsis thaliana*, *Sci Total Environ* (461-462) : 462–468
178. Savithramma N, Ankanna S, Bhumi G, (2012) Effect of nanoparticles on seed germination and seedling growth of *Boswellia ovalifoliolata* an endemic and endangered medicinal tree taxon. *Nano Vision* 2:61–68.
179. Krishnaraj C, Jagan EG, Ramachandran R, Abirami SM, Mohan N, Kalachelvan PT (2012) Effect of biologically synthesized silver nanoparticles on

- Bacopa monnieri* (Linn.) Wettst. Plant growth metabolism. *Process Biochem* 47(4): 651–658
180. N. Gruyer, M. Dorais, C. Bastien, N Dassylva, G. Triffault-Bouchet (2013) Interaction between silver nanoparticles and plant growth. *Atta-Hortic*. 1037 : 795-800
181. Yang F, Liu C, Gao F, Su M, Wu X, Zheng L et al (2007) The improvement of spinach growth by nano-anatase TiO₂ treatment is related to nitrogen photoreduction. *Biol.Trace Elem. Res.* 119: 77–88.
182. Song G, Gao Y, Wu H, Hou W, Zhang C, Ma H (2012) Physiological effect of anatase TiO₂ nanoparticles on *Lemna minor*. *Environ. Toxicol. Chem.* 31:2147–2152
183. Zheng L, Hong F, Lu S, Liu C, (2005) Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biol.TraceElem.Res.*105:83–91.
184. E.H. Dehkourdi, M. Mosavi (2013) Effect of anatase nanoparticles (TiO₂) on parsley seed germination (*Petroselinum crispum*) in vitro *Biol.TraceElem.Res.* 155 (2): 283–286
185. Feizi H, Kamali M, Jafari L, Rezvani Moghaddam P (2013) Phytotoxicity and stimulatory impacts of nanosized and bulk titanium dioxide on fennel (*Foeniculum vulgare* Mill). *Chemosphere* 91(4):506–511.
186. Lu CM, Zhang CY, Wen JQ, Wu GR, Tao MX (2002) Research of the effect of nanometer materials on germination and growth enhancement of *Glycine max* and its mechanism. *Soybean Sci.* 21: 168–172.
187. Zheng L, Hong F, Lu S, Liu C (2005) Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biol Trace Elem Res.*104:83–91.
188. Hong F, Zhou J, Liu C, Yang F, Wu C, Zheng L, Yang P (2005) Effect of nano-TiO₂ on photochemical reaction of chloroplasts of spinach. *Biol Trace Elem Res*, 105: 269–279.
189. Lopez-Moreno ML, De La Rosa G, Hernandez-Viezcás JA, Castillo-Michel H, Botez CE, Peralta-Videa JR, Gardea-Torresdey JL (2010) Evidence of the differential biotransformation and genotoxicity of ZnO and CeO₂ nanoparticles on soybean (*Glycine max*) plants. *Environ Sci Technol* 44:7315–7320.
190. H. Wang, X. Kou, Z. Pei, J.Q. Xiao, X. Shan, B. Xing (2011) Physiological effects of magnetite (Fe₃O₄) nanoparticles on perennial ryegrass (*Lolium perenne* L.) and pumpkin (*Cucurbita mixta*) plants, *Nanotoxicology*. 10.3109/17435390.489206.5(1): 30-42
191. Sheykhbaglou R, Sedghi M, Shishevan MT, Sharifi RS (2010) Effects of nano-iron oxide particles on agronomic traits of soybean. *Notulae Scientia Biologicae* 2: 112–113.
192. T N V K V. Prasad, P. Sudhakar, Y Sreenivasulu, P. Latha, V. Munaswamy, K.R. Reddy (2012) Effect of Nanoscale Zinc oxide particle on the Germination, Growth and yield of Peanut *J.PlantNutr.* 35 (6) : 905–927.
193. Suriyaprabha R, Karunakaran G, Yuvakkumar R, Rajendran V, Kannan N (2012) Silica nanoparticles for increased silica availability in maize (*Zea mays* L) seeds under hydroponic conditions. *Curr Nanosci* 8: 902–908.

194. Jo YK, Kim BH, Jung G (2009) Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Dis* 93(10):1037–1043.
195. Lamsal K, Kim SW, Jung JH, Kim YS, Kim KS, Lee YS (2011) Application of silver nanoparticles for the control of *Colletotrichum* species in vitro and pepper anthracnose disease in field. *Mycobiology* 39(3):194–199.
196. Gajbhiye M, Kesharwani J, Ingle A, Gade A, Rai M (2009) Fungus-mediated synthesis of silver nanoparticles and their activity against pathogenic fungi in combination with fluconazole. *Nanomed-Nanotechnol* 5(4):382–386.
197. C.O. Dimkpa, J.E. McLean, D.W. Britt, A.J. Anderson (2013) Antifungal activity of ZnO nanoparticles and their interactive effect with a biocontrol bacterium on growth antagonism of the plant pathogen *Fusarium graminearum*, *Biometals* 26(6) : 913-24
198. Huang L, Li DQ, Lin YJ, Wei M, Evans DG, Duan X (2005) Controllable preparation of nano-MgO and investigation of its bactericidal properties. *J Inorg Biochem* 99(5):986–993
199. Kanhed P et al (2014) In vitro antifungal efficacy of copper nanoparticles against selected crop pathogenic fungi. *Mater Lett* 115:13–17.
200. Jo YK, Kim, B. H., Jung G H (2009) Antifungal activity of silver ions and nanoparticles on phytopathogenic fungi. *Plant Disease* 93:1037-1043.
201. Kasprowicz M J, Koziół M, Gorczyca A (2010) The effect of silver nanoparticles on phytopathogenic spores of *Fusarium culmorum*. *Canadian Journal of Microbiology* 56: 247-253.
202. Lamsal K, Sang-Woo Kim¹, Jin Hee Jung¹, Yun Seok Kim¹, Kyoung Su Kim² Youn Su Lee (2010) Inhibition Effects of Silver Nanoparticles against Powdery Mildews on Cucumber and Pumpkin. *Microbiology* 39(1):26-32
203. Gopinath K, Karthika V, Sundaravadivelan C, Gowri S, Arumugam A (2015) Mycogenesis of cerium oxide nanoparticles using *Aspergillus niger* culture filtrate and their applications for antibacterial and larvicidal activities. *Journal of nanostructure in chemistry* 5:295–303
204. Ghormade V, Deshpande MV, Paknikar KM (2011) Perspectives for nanobiotechnology enabled protection and nutrition of plants. *Biotechnol. Adv* 29:792-803.
205. Gruère G, Clare N, Linda A (2011) Agricultural, Food and Water Nanotechnologies for the Poor Opportunities, Constraints and Role of the Consultative Group on International Agricultural Research. *J. Int. Food Policy Res. Inst* 1-35.
206. Nair R, Varghese SH, Nair BG, Maekawa T, Yoshida Y, Kumar DS, Nanoparticulate material delivery to plants. *Plant Sci* 179:154-163.
207. Naderi MR, Danesh-Shahraki A. (2013) Nanofertilizers and their roles in sustainable agriculture. *Int J Agri Crop Sci* 5(19):2229–2232.
208. Moaveni P, Kheiri T (2011) 2nd International Conference on Agricultural and Animal Science; November 25–27, 2011; Maldives. Singapore: IACSIT Press (2011) 22:160–163.

209. Liu R, Lal R, (2014) Synthetic apatite nanoparticles as a phosphorus fertilizer for soybean (*Glycine max*), *Sci. Rep.* 4:5686
210. Shah V & Belozerovala I (2009) Influence of metal nanoparticles on the soil microbial community and germination of lettuce seeds. *Water Air Soil Pollut* 197:143–148.
211. Ambrogio MW, Frascioni M, Yilmaz MD, Chen X (2013) New methods for improved characterization of silica nanoparticlebased drug delivery systems. *Langmuir* 29:15386–15393.
212. Ghafariyan MH, Malakouti MJ, Dadpour MR, Stroeve P, Mahmoudi M (2013) Effects of magnetite nanoparticles on soybean chlorophyll. *Environ. Sci. Technol* 47:10645–10652.
213. Torney F, Trewyn BG, Lin VS, Wang K (2007) Mesoporous silica nanoparticles deliver DNA and chemicals into plants. *Nat. Nanotechnol* 2:295–300.
214. First nano-organic iron chelated fertilizer invented in Iran. Iran Nanotechnology Initiative Council; (2009) :http://www.iranreview.org/content/Documents/Iranians_Researchers_Produce_Nano_Organic_Fertilizer.html. Accessed April 11, 2014.
215. Iranian scientist develops nano organic fertilizer (2014) Meher news agency <https://en.mehrnews.com/news/1004486/Iranian-scientist-Nano-organic-Fertilizer>. Accessed October 29, 2014.
216. Manikandan A, Subramanian KS (2014) Fabrication and characterisation of nanoporous zeolite based N fertilizer. *Afr J Agric Res* 9(2):276–284.
217. El-Ramady HR (2014) Integrated Nutrient Management and Postharvest of Crops. *Sustainable Agri Rev* 13:163–274.
218. López MM, Llop P, Olmos A, Marco-Noales E, Cambra M, Bertolini E (2009) Are molecular tools solving the challenges posed by detection of plant pathogenic bacteria and viruses? *Curr. Issues Mol. Biol.* 11(1):13-46
219. Yao KS, Li SJ, Tzeng KC, Cheng TC, Chang CY, Chiu CY, Liao CY, Hsu J, Lin ZP (2009) Fluorescence silica nanoprobe as a biomarker for rapid detection of plant pathogens. *Adv. Mater. Res* 79(82):513-516

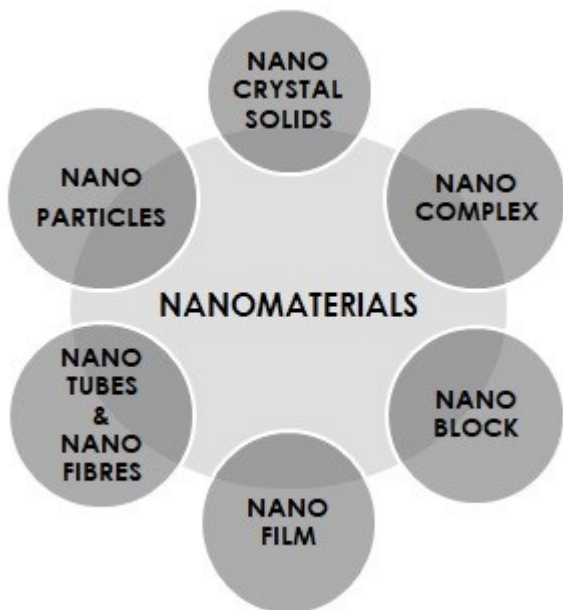


Figure1. Classification of nanomaterials

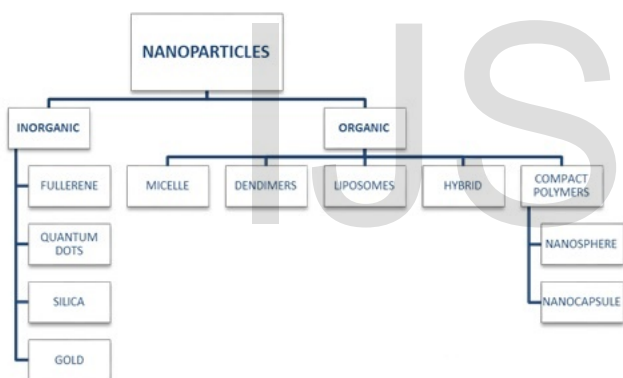


Figure2. Different Types of nanoparticles.

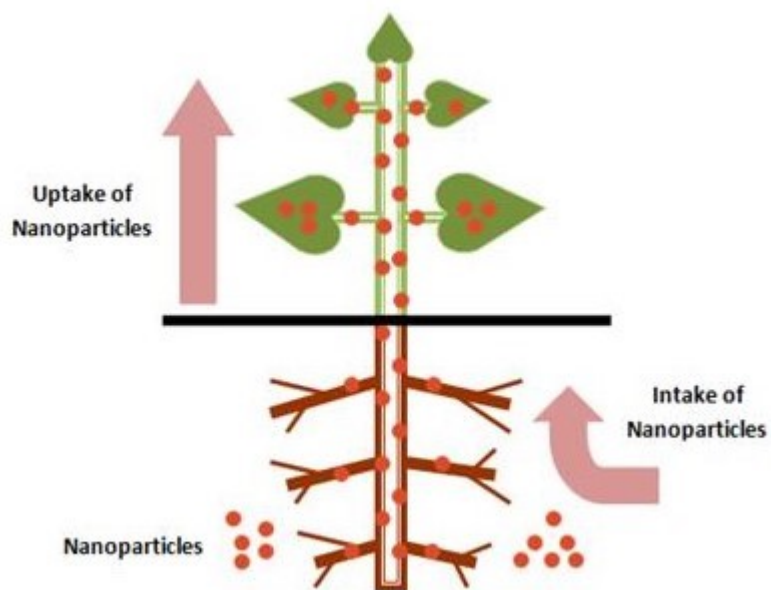


Figure3. A cartoon depicting the entry of nanoparticles in the plant system.

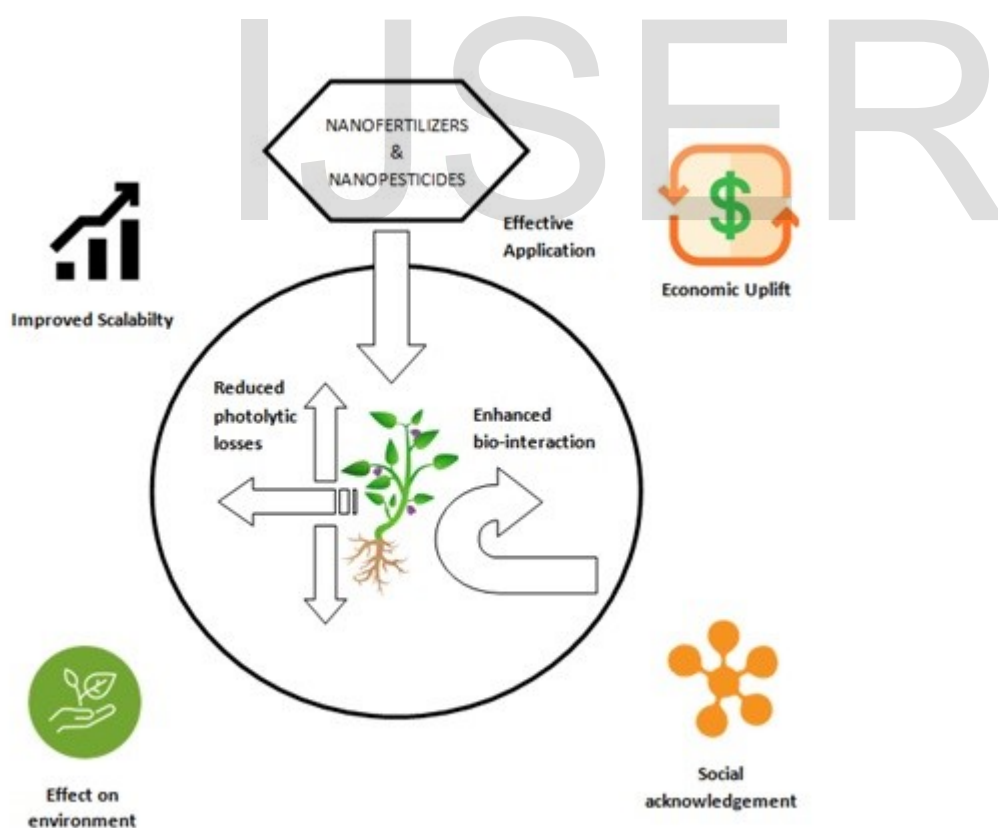


Figure4. A cartoon depicting efficacy of Nanofertilizers and Nanopesticides

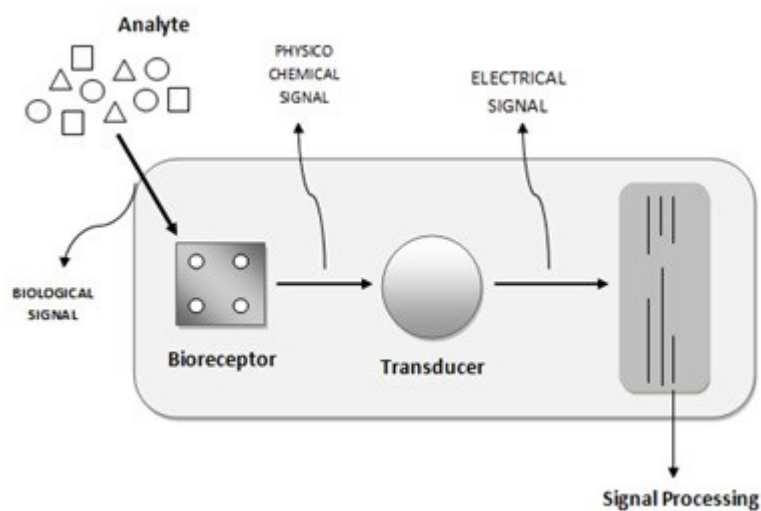


Figure5. A picture depicting the mechanism of biosensor.

IJSER